



Neurofibromatosis Type 1: Bridging Science and Patient-Centered Care

Kiyoharu J. Miyagishima ¹, Fengyu Qiao ¹, Steven F. Stasheff ^{1,2,3} and Francisco M. Nadal-Nicolás ^{1,*}

- Retinal Neurophysiology Section, National Eye Institute, National Institutes of Health, Bethesda, MD 20892, USA; kiyoharu.miyagishima@nih.gov (K.J.M.); fengyu.qiao@nih.gov (F.Q.); sfstasheff@childrensnational.org (S.F.S.)
- ² Center for Neuroscience and Behavioral Medicine, Gilbert Neurofibromatosis Institute, Children's National Health System, Washington, DC 20010, USA
- ³ Neurology Department, George Washington University School of Medicine, Washington, DC 20037, USA
- * Correspondence: nadalnicolasfm@nih.gov

Abstract: Neurofibromatosis type 1 (NF1) is an inherited autosomal dominant disorder primarily affecting children and adolescents characterized by multisystemic clinical manifestations. Mutations in neurofibromin, the protein encoded by the *Nf1* tumor suppressor gene, result in dysregulation of the RAS/MAPK pathway leading to uncontrolled cell growth and migration. Neurofibromin is highly expressed in several cell lineages including melanocytes, glial cells, neurons, and Schwann cells. Individuals with NF1 possess a genetic predisposition to central nervous system neoplasms, particularly gliomas affecting the visual pathway, known as optic pathway gliomas (OPGs). While OPGs are typically asymptomatic and benign, they can induce visual impairment in some patients. This review provides insight into the spectrum and visual outcomes of NF1, current diagnostic techniques and therapeutic interventions, and explores the influence of NF1-OPGS on visual abnormalities. We focus on recent advancements in preclinical animal models to elucidate the underlying mechanisms of NF1 pathology and therapies targeting NF1-OPGs. Overall, our review highlights the involvement of retinal ganglion cell dysfunction and degeneration in NF1 disease, and the need for further research to transform scientific laboratory discoveries to improved patient outcomes.

check for updates

Citation: Miyagishima, K.J.; Qiao, F.; Stasheff, S.F.; Nadal-Nicolás, F.M. Visual Deficits and Diagnostic and Therapeutic Strategies for Neurofibromatosis Type 1: Bridging Science and Patient-Centered Care. *Vision* **2024**, *8*, 31. https://doi.org/ 10.3390/vision8020031

Received: 2 March 2024 Revised: 3 May 2024 Accepted: 4 May 2024 Published: 9 May 2024



Copyright: © 2024 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). **Keywords:** NF1; pediatric low-grade glioma; childhood; optic nerve; chiasm; ocular pathologies; animal models; mice; OCT; chemotherapy

1. Introduction

Neurofibromatosis Type 1 (NF1) is a rare, multifaceted genetic disorder with a complex spectrum of phenotypic clinical manifestations making treatment challenging. The most common feature is café au lait patches on the skin [1]. The prevalence of NF1 is reported to be one in ~3500 individuals [2,3], and although NF1 is an autosomal dominant condition, ~50% of cases occur by *de novo* mutations [4]. NF1 stems from mutations in the *Nf1* tumor suppressor gene, located on chromosome 17q11.2, encoding the neurofibromin protein [5]. Neurofibromin regulates the activity of the RAS-MAPK signaling pathway crucial for cell growth and division [6,7]. In normal conditions, neurofibromin binds RAS to regulate RAF-MEK-ERK activation of the MAPK pathway; however, mutations in the *Nf1* gene result in reduced or absent neurofibromin activity in individuals with NF1, causing uncontrolled cell growth and tumor formation (Figure 1) [8]. Neurological manifestations begin at birth or during early childhood. Individuals are at increased risk of developing tumors of the central nervous system (CNS), including the brain and spinal cord [9]. Clinical manifestations can include benign peripheral nerve sheath tumors, bone deformation, and even curvature of the spine (scoliosis). Scoliosis, which may affect motor abilities,

is estimated to be present in ~20% of children with NF1, accounting for approximately ~2% of all pediatric scoliosis cases [10,11]. Features that may also be present include short stature and macrocephaly [12]. Although most NF1 individuals display normal intelligence, learning disabilities are quite common [13–16].



Figure 1. Diagram illustrating the formation of optic pathway gliomas (OPGs) in children with Neurofibromatosis Type 1. Under normal conditions, growth factors stimulate the activation of RAS-GDP to RAS-GTP through son of sevenless (SOS). Neurofibromin (*Nf1* gene) regulates the conversion of RAS-GTP to its inactive form (RAS-GDP), thereby modulating cell growth and migration through the MAPK/ERK and mTOR pathways. In NF1 patients, mutations of neurofibromin significantly reduce its natural activity, resulting in abnormal hyperactivation of the MAPK/ERK and mTOR pathways. Consequently, uncontrolled cell growth and migration lead to the development of gliomas in the optic pathway, potentially affecting vision. Created with BioRender.com.

This review specifically focuses on visual abnormalities associated with NF1. Tumors of the optic nerve associated with NF1, termed optic pathway gliomas (OPGs), are usually benign. OPGs can occur anywhere along the optic pathway, from the optic nerve to the optic chiasm, and they can cause visual disturbances that can be either anatomical (strabismuseye misalignment) or functional (by decreasing the visual field or visual acuity) [17]. However, some NF1 individuals experience visual deficits that cannot be fully explained by the presence of OPGs. In light of this, there is evidence from animal models suggesting that individuals with NF1 may be at higher risk for retinal ganglion cell (RGC) dysfunction and degeneration [18]. RGCs are a unique neuronal cell type located in the innermost portion of the retina that transmit visual information along their axons (forming the optic nerves) to the brain [19,20]. RGCs are a vital component for visual function and other non-imageforming functions such as circadian photoentrainment and the pupillary reflex [21,22]). Like other neurons in the CNS [23] which lack regenerative capacity [24], RGC injury often leads to cell death and permanent vision loss.

The mechanism underlying RGC degeneration in NF1 involves alterations in genes and signaling pathways that regulate cell growth, differentiation, and survival. Further research is needed to fully understand the relationship between NF1 and RGC degeneration, and to develop effective treatments for this condition. Although some individuals with NF1 have a distinct genotype/phenotype correlation [25,26], heterogeneity in clinical presentation is observed in patients and could be attributed to stochastic events, environmental factors, or modifier genes [27–29]. Advances in imaging now allow for non-invasive examination of the retina (optical coherence tomography, OCT) and tumor size and position (magnetic resonance imaging, MRI; magnetic resonance spectroscopy, MRS), offering invaluable "in vivo" measurements. NF1 cases can be examined by infrared fundus autofluorescence (IR-FAF) and OCT to characterize choroidal abnormalities [30] in addition to measurements of retinal nerve fiber layer thickness [31]. Using magnetic resonance imaging (MRI) with contrast enhancement, the optic nerve sheath complex in patients with optic pathway gliomas can be visualized as hypointense on T1-weighted images and hyperintense on T2-weighted images compared to the normal optic nerve [32,33]. Measured parameters include diameter and signal intensity of the optic nerve as well as degree of tortuosity [34]. For diagnostic purposes, imaging and clinical examination is sufficient; however, it is difficult to link these observations to predict vision loss attributed to retinal ganglion cell loss or tumor progression. Ex vivo examination of OPGs is rare as surgical removal is an uncommon practice [35,36]. Thus, the scarcity of enucleated eyes in pediatric patients prevents drawing correlations between RGC quantification, which can only be performed ex vivo, with measurable parameters in patients such as tumor size, type, position, and degree of associated visual impairments. Currently, image analysis and machine learning [37] are being applied to MRIs of optic pathway gliomas to build predictive models that may one day compliment ex vivo methodologies for tumor classification [38,39] or RGC quantification [40,41]. This comprehensive review focuses on neuronal tissues and explores the ocular symptoms associated with NF1, linking current research with advances in diagnostic and therapeutic strategies.

2. Phenotypic Manifestations of NF1 Affecting Vision

NF1 ocular manifestations exhibit significant clinical heterogeneity during childhood and adolescence. From overt signs such us Lisch nodules (LNs) to complex ophthalmological complications (such as optic gliomas, plexiform neurofibromas, and congenital glaucoma), there is a full spectrum of pathologies that can influence visual acuity and perceived visual field [42]. LNs, benign tumors with a yellowish-brown dome shape that grow over the iris surface, tend to increase in size and number with age [43,44]. Vision impairment in NF1 patients may stem from various factors, including anatomical causes like proptosis or strabismus originating from intraorbital and periorbital (eye lid and face) plexiform neurofibromas (PLXNs) [17] that may obstruct vision, displace the location of the eye, and interfere with ocular motility; sphenoid wing dysplasia; and the presence of tumors along the optic nerve, OPGs [45–47].

OPGs are predominantly asymptomatic low-grade gliomas (LGGs), primarily affecting the anterior visual pathway, with 75% occurring in the optic nerve (ON) and optic chiasm. However, they can often involve both ONs, posterior visual pathway segments (optic tract and radiations), and the hypothalamus [48,49]. While mainly pilocytic astrocytomas (grade I), OPGs can also be pilomyxoid astrocytomas and diffuse fibrillary astrocytomas (grade II) [50,51], exhibiting clinical variability. Depending on their behavior, OPGs can be aggressive, leading to visual loss [49,52,53], or regress spontaneously [54]. Symptomatic gliomas typically manifest before age 6, with an average onset of 4.5 years [55]. The location, type, size, and number of OPGs can cause various neurological symptoms affecting visual function and resulting in a range of visual deficits (Figure 2) [56,57]. OPGs may occur unilaterally or bilaterally, be situated anterior to, posterior to, or at the chiasm, centered or asymmetric. The optic tract and even the hypothalamus can also be involved (Figure 2) [53,58]. In extreme cases OPGs can also reach the lateral geniculate nuclei and

temporal lobes [59]. Larger tumors and those located closer to the optic nerve are usually associated with more severe visual impairment. Interestingly, patients with gliomas isolated to the optic nerve have better long-term visual outcomes than those with postchiasmatic involvement [60,61]. Additionally, LGGs can affect other brain areas, known as non-OPGs [62]. Although non-OPGs are less frequent than OPGs, they are more frequent in older children/young adults [51] and they can also cause a wide range of neurological symptoms depending on their location, size, and number. These symptoms include headaches, seizures, changes in behavior, cognitive problems, or visual deficits if higher areas of visual processing are involved [63].

Beyond the visual pathway, advancements in multimodal imaging in ophthalmology have revealed microvascular abnormalities in the retinas of NF1 patients [64,65], potentially causing progressive insults of ischemic injury that affect RGC function and viability. Additionally, choroidal abnormalities and hyperpigmented spots have also been observed during ophthalmologic examinations [66]. Although the effects of these choroidal abnormalities on vision are not established yet, the choroid plays a crucial role in maintaining the retinal pigmented epithelium (RPE) and the photoreceptors. Given that photoreceptors are highly metabolically active with high oxygen consumption, choroidal abnormalities impacting oxygen delivery may adversely affect their survival and function and have a profound effect on the patient's vision. Thus, the diverse ocular manifestations in NF1 individuals highlight the significance of early diagnosis in improving clinical management and enhancing patient outcomes.



Figure 2. Visual field defects based on the location and size of the axonal damage in the optic pathway. (**A**) Schematic representation of the retinal ganglion cell (RGC) axon projection to superior brain areas for visual processing in the brain. Ipsilaterally (blue) or contralaterally (black and red) projecting RGCs within the optic nerves. (**B**) Depiction of individualized visual field deficits in patients with axonal damage in the optic pathway corresponding to their respective scheme in which red marks represent OPG size and location. Drawings based on concepts presented in [56,57,67].

Although OPGs are a classic characteristic of NF1 pathology, sporadic OPGs (not associated with NF1) can also form exhibiting distinct genetic hallmarks. In sporadic OPGs, the most common genetic alteration is a duplication of the kinase domain of a gene called B-Raf (BRAF) that leads to MAPK pathway activation, promoting cell survival, growth, and proliferation [68–70]. However, other genetic mutations in sporadic OPGs can occur in KRAS, RAF1, FGFR1, PTPN11, and NTRK2 genes [71–74]. Although NF1 associated OPGs are predominantly found in females, sporadic OPGs occur in both genders with similar frequency [50,75]. Like NF1, sporadic OPGs are often symptomatic and clinically more severe, with aggressive tumor growth and rapid visual decline compared to OPGs associated with

5 of 19

NF1 [75–79]. Notably, studies have determined that sporadic OPGs are predominantly located at the optic chiasm, while OPGs associated with NF1 are generated at the chiasm as well as the optic nerve [76,80–82]. Despite their location, sporadic OPGs can lead to a large variety of symptoms such as proptosis, nystagmus, hypothalamic-related endocrine alterations, hydrocephalus, raised intracranial pressure, and vision loss [81,83]. In fact, sporadic OPGs carry a higher risk of vision loss (66–74% in comparison with a 50% risk in NF1-associated OPGs), with visual deficiencies that can appear bilaterally in 25% of cases, and progressive visual loss in 74% of patients regardless of therapy [77,84]. Similar to NF1 OPGs, treatment is necessary only if accompanied by visual impairment. The main treatment is chemotherapy which will be discussed in further detail below.

3. Diagnosis and Monitoring Methodologies

The diagnosis of NF1 primarily relies on clinical criteria established by the National Institutes of Health (NIH) in 1987 [85]. These criteria are based on characteristic features such as café au lait spots and neurofibromas. Thus, children meeting clinical criteria for NF1 are expected to undergo regular eye exams to detect asymptomatic OPGs. In fact, several studies have reported a high prevalence of brain tumors in asymptomatic children [44]. Similarly, children presenting with unexplained vision loss, monocular or asymmetric nystagmus, or optic atrophy should be considered for NF1 clinical diagnosis [86]. However, numerous children present significant vision loss prior to receiving treatment, emphasizing the importance of early detection and intervention [87]. Therefore, conducting annual screenings and longitudinal monitoring for signs and symptoms related to OPG throughout childhood is essential for timely clinical decision-making and treatment [88].

In terms of vision, Lisch nodules (LNs) serve as pathognomonic markers of NF1, indicating a potential vision-threatening condition [55]. They appear as small dome-shaped lesions on the iris of the eye and although LNs do not cause visual disturbances, their presence is rare in individuals without NF1. Detecting LNs through a slit lamp examination is a straightforward, noninvasive, and cost-effective method for accurately diagnosing NF1. However, the absence of LNs at earlier stages does not exclude NF1 diagnosis [44]. Importantly, there is no established association between LN presence and the overall clinical severity of NF1 pathology in patients [89,90].

Traditionally, biopsies have been the standard investigation method to confirm NF1 tumor diagnosis. However, they are no longer used routinely, but only exceptionally, to confirm the presence of OPGs [91]. Several imaging techniques offer non-invasive examination of OPGs. Advanced imaging techniques, such as computed tomography (CT) and magnetic resonance imaging (MRI), are ideal to visualize the entire optic nerve or the optic pathway. However, MRI is the preferred method because of superior soft tissue resolution [92–94], since OPGs cannot be easily detected on CT scans. Non-contrast MRI provides accurate measurements of tumor size, and contrast-enhanced MRI precisely delineates involvement of other adjacent areas, such as the hypothalamus [94]. MRI can be used to monitor tumor progression in NF1 patients using different approaches, such as volumetric analysis or linear measurements [95]. However, radiographic results to date have provided poor correlation with functional outcomes of patients (visual acuity) in numerous studies [92,93,96].

Interestingly, machine learning algorithms can aid in analyzing the large amount of data generated by these techniques, facilitating initial screening for doctors [97]. Notably, a study reported a correlation between the OPG volume and RGC axon loss [98]. However, systematic MRI screening in children with NF1 has not shown clear benefits yet [99], as early OPG diagnosis and treatment did not improve visual outcomes. Without follow-up examinations, continued optic glioma growth cannot be excluded [8,100]; thus, annual ophthalmologic assessment for changes in the patient's vision is always recommended.

Optical coherence tomography (OCT) uses reflected near infrared light to produce cross sectional images of retinal tissue structure with a depth of several hundred microns and can help in the diagnosis and evaluation of OPG-associated pathology. In NF1 patients,

RGC degeneration can be evaluated by measuring the thickness of the different retinal layers [101]. The ganglion cell complex (GGC) and retinal nerve fiber layer (RNFL [102,103]) thickness provide a quantitative measure of RGC viability. The GCC, comprising the ganglion cell layer and inner plexiform layer, reflects the status of the RGC somas and dendrites, excluding their axons. The RNFL, on the other hand, is primarily composed of RGC axons that converge at the optic nerve head to form the optic nerve. Importantly, the RNFL also includes processes from glial cells (astrocytes, microglia, and Müller cells) that form an intricate interlocking pattern with RGC axons in primate retinas [104,105]. Thus, reactive and infiltrating glial cells in pathological conditions may impact measurements of RNFL thickness, complicating interpretation. Unfortunately, the use of different imaging instrumentation and proprietary software have made it difficult to make direct comparisons among studies [106,107]. In addition, studies in rodents suggest a potential mismatch between RGC loss and RNFL thinning, further complicating interpretation [104,108]. These experiments indicate RGC death precedes axonal atrophy and removal; however, the experimental insult of axotomizing the RGC axons may not recapitulate OPG damage. Despite these caveats, significant thinning of the RNFL or the GCC implies RGC loss, and patients may show substantial visual impairment. Thus, although measurements of RNFL or GCC thickness do not provide a causative link to visual outcomes, they can identify associated anatomical changes to the retina.

Near-infrared imaging (NIR) of the fundus (rear of the eye) generates a 2D image from the amount of reflected light. NIR is able to detect choroidal nodules in NF-1 patients, which appear bright (hyperreflective) and patchy [109]. Although these choroidal nodules do not affect RGCs and their axons directly, they may affect epithelial transport functions of the RPE between the choroid and retina, potentially affecting photoreceptor viability and visual function. Nonetheless, evaluating visual acuity may not identify minor photoreceptor loss, as numerous studies have indicated that a 40–60% loss of cone photoreceptors in the fovea does not have an impact on visual acuity [110–112].

Functional tests, particularly visual evoked potentials (VEPs), might play a crucial role in diagnosing and monitoring progression. In response to visual stimulation, VEPs record the generation of electrical impulses from the visual cortex in the brain through electrodes placed in the scalp [31,113]. Reduced amplitudes or delayed responses indicate the magnitude of visual deficits; however, they cannot reveal the nature of the vision loss, and some reports question the correlation between VEP evaluations and vision loss [114–116].

A potential future ideal would be molecular and genetic testing to predict the likely phenotype and complications for a person with a specific *Nf1* germline mutation. However, genetic testing is rarely performed due to the extensive heterogeneity in the mutations of the neurofibromin gene and the potential influence of stochastic factors. To date, over 3000 pathological genetic variants of the *Nf1* gene have been identified [117], with less than 20% reported as recurrent [118,119]. Despite the majority of investigations not establishing genotype–phenotype correlations or providing inconclusive results [53], recent studies employing novel screening techniques are beginning to establish correlations [25,120–129]. The integration of genetic testing in future studies could play a pivotal role in confirming NF1 diagnoses and uncovering genetic variations that may influence ocular phenotypes. Thus, compiling a library of cases has the potential to diagnose and classify new patients earlier, facilitating treatment decision making.

Currently, NF1 gene mutation testing is performed primarily to help confirm or reduce the likelihood of an NF1 diagnosis in cases of clinical uncertainty [86], though genetic testing can also be expanded to include family testing in relatives with uncertain clinical signs or for prenatal testing [6].

Early diagnosis of NF1 patients could be challenging but it is crucial for successful intervention [55,63]. The abovementioned examinations require patient cooperation that can be complicated in preverbal children, especially if they are cognitively compromised. In addition, the natural history of OPGs in NF1 is highly variable; some tumors remain stable, others can regress over time, while others progress, causing significant vision

loss. However, these techniques provide crucial information to guide treatment decision, although decisions remain complex [91,130]. Thus, monitoring is an integral part of the management of NF1-associated vision impairment, and examples such as positron emission tomography have proven useful for monitoring OPG progression and response to treatments [131].

4. Therapeutic Strategies

Treatment strategies for children with NF1-related visual complications attempt to halt progressive vision loss and promote healthy development. OPG treatment approaches vary depending on specific symptoms, tumor location and size, and extent of visual impairment, and may involve observation, surgical procedures, radiation, chemotherapy, or targeted therapies.

The presence of LNs rarely interfere with vision and typically do not require treatment [44]. However, when OPGs result in the compression of the optic nerve, surgery can reduce symptoms. Surgical procedures are seldom performed to remove pediatric LGGs because they are benign and the likelihood of progression to the chiasm or potential damage to the fibers crossing from the contralateral eye is low [35,36]. Intraorbital and especially intracranial procedures are invasive and carry the risk of vision loss and they can be potentially life-threatening [132], due to bleeding complications [133,134]. Thus, surgical procedures are warranted based on anatomical location and accessibility [135]. Surgery is primarily recommended in cases involving pain, disfiguring proptosis, and/or compression of the surrounding tissues [136]. Although surgery is unlikely to improve vision in patients with orbital OPGs, it may be undertaken for cosmetic purposes [57] or for biopsy if the eye is blind.

Radiotherapy offers an effective treatment for OPGs, but its use is mainly limited to teenagers and those without targeted treatment options due to potential adverse effects, especially in young patients with developing brains [136]. These adverse effects include reduced visual function [137,138], neurocognitive deficits [138–140], cerebrovascular abnormalities [141,142], and alterations in endocrine function [143,144] that can persist into adulthood. Novel options include 3D conformal radiation therapy where radiation beams are matched to the volumetric shape of the cancer [145], stereotactic radiosurgery (gamma knife) that focuses the beam to treat smaller targets [146,147], and fractionated stereotactic radiation [148,149] and proton beam radiation [59,150] which minimize damage to healthy surrounding tissue.

Currently, OPGs displaying substantial progression are treated with chemotherapy [47,136,151]. Vincristine and carboplatin are often prescribed as the first line of treatment and have shown reasonable progression-free survival rates at earlier stages [152]. However, carboplatin doses can lead to hypersensitivity and frequency-based adverse effects in some individuals [153]. Alternative combinations, such as cisplatin and etoposide [154] or thioguanine, procarbazine, and lomustine appear to improve event-free survival [155]. However, caution on the use of these alternative drugs in treating NF1 is advised due to the risk of developing secondary leukemia attributed to etoposide [156], and to procarbazine and lomustine [154,157]. Monotherapies with vinblastine [158,159], vinorelbine [160], trametinib [135,161] or temozolomide [162] have also shown efficacy in NF1 patients with low toxicity, except there have been reported cases of secondary leukemia following temozolomide and radiotherapy [163]. Notably, pre-chiasmatic OPGs appear to be more responsive to chemotherapy than gliomas located elsewhere in the optic pathway [164]. However, while chemotherapy often limits tumor growth effectively, very few individuals show visual improvement following treatment [96], particularly patients with late-progressive OPGs [165].

Another set of treatments are targeted therapies. Bevacizumab is an anti-vascular endothelial growth factor (VEGF) monoclonal antibody [166] that reduces vascular permeability and tumor growth. It improves visual symptoms in most cases [167,168] when administrated alone or in combination with irinotecan, a DNA topoisomerase I inhibitor that interrupts DNA replication and cancer growth [169–172]. However, bevacizumab causes reversible side effects (hypertension, fatigue, joint pain, bleeding, and proteinuria) that can persist after treatment ending, and tumor progression is common after treatment discontinuation [167,171,173].

Novel agents focus on inhibiting the mitogen-activated protein kinase (MAPK) pathways. Selumetinib, a selective MEK1/2 inhibitor, has been shown to maintain or improve visual acuity after oral administration in patients with OPGs [174]. Recent studies show MEK inhibitors such as refametinib, trametinib, and cobimetinib can shrink the volume of most inoperable benign LGGs and malignant plexiform neurofibromas, improving neurocognitive function in NF1 patients [175,176]. In fact, MEK inhibitors have also demonstrated tumor suppression in preclinical mouse models [177]. However, it is important to note that the response of LGGs to MEK inhibition is often variable, and regrowth is frequently observed after discontinuation of therapy [177]. Other promising treatment options include small competitive molecules (vemurafenib and dabrafenib) that prevent bRAF from binding MEK and activating the MAPK pathway [178]. Rapamycin and its derivates, such as everolimus, are selective mTOR blockers. A recent study demonstrated oral administration of everolimus stabilized visual acuity in children with NF1-OPGs with low levels of toxicity [179,180].

In addition, administration of pro-survival factors can preserve RGC survival or stimulate axonal regrowth, as demonstrated by a clinical study in which NF1 patients were given eye drops with murine nerve growth factor for 10 days, and a third of the treated group showed significant improvement in the size of their visual field [181].

Overall, the management of NF1-associated vision impairment requires a multidisciplinary approach, involving close collaboration between ophthalmologists, oncologists, neurologists, geneticists, neurosurgeons, endocrinologists, and pathologists. Early diagnosis is crucial to initiate treatment promptly and prevent irreversible visual decline, while regular monitoring (tumor size, visual function) is essential to ensure optimal outcomes in patients with NF1 and OPGs.

5. The Role of Animal Models to Uncover Underlying Mechanisms of NF1 and to Develop Novel Therapies

The scarcity of surgical resection or biopsies from OPGs in NF1 patients underscores the utility of preclinical animal models in providing knowledge about these tumors.

Genetically engineered rodents, particularly mice, are the most widely used and best characterized models of NF1-OPG. Generation of Nf1 knockout mice from the germline $(Nf1^{-/-})$ was unsuccessful as they were lethal and their heterozygous littermates $(Nf1^{+/-})$ did not develop astrocytomas despite increased astrocyte proliferation [182,183]. Subsequent studies focused on germline mutations in Nf1 resulting in varying levels of neurofibromin expression and the development of optic gliomas [184]. Currently, the most successful mouse models are the conditional knockout lines, which allow for inactivation of Nf1 in specific cell lineages. Utilizing the Cre-lox system enables the generation of NF1-associated tumors in animals without being lethal. In this context, specific deletion of the Nf1 gene in astrocytes (GFAP-Cre; Nf1^{flox/mut}) successfully induced formation of OPGs [185,186]. Additional mouse lines have been developed that more closely resemble NF1 human pathology by introducing human GFAP (hGFAP-Cre; Nf1^{flox/mut}) [187]. This line exhibited complete penetrance of glial hyperplasia and enlarged optic nerves with lesions that in some cases progressed to form optic pathway gliomas [188]. Other mouse models achieve OPG formation by activating the KRAS oncogene in astrocytes of heterozygous Nf1 mice [189], or inhibiting Nf1 in neuroglial progenitors (such as BLBP and Oligo2 [190,191]) thereby increasing proliferation of cells with glial lineage and inducing abnormal neuronal differentiation. However, a recent report suggests that in Nf1-deficient neuroglial progenitor cells, CNS injury could be sufficient to induce glioma formation, indicating that independent injuries can promote tumor development in susceptible animals [192]. Mouse models have contributed to our understanding of OPG formation and have highlighted mechanisms for mTOR-dependent glioma formation [193], implicating microglia in glioma formation [194–197], and the presence of glioma-specific stem cells [198,199]. In addition, mouse models are a valuable tool to design novel therapeutic strategies or redefine existing treatments. Animal studies have helped define the therapeutic window to rescue neural progenitors by administration of MEK/ERK inhibitors during early postnatal stages [200]. Interestingly, Jecrois and colleagues reported that either the simultaneous removal of three out of the four alleles from the Mek1 and Mek2 genes (as complete elimination of the two alleles of Mek1 and the two alleles of Mek2 proved lethal) or administration of a low-dose MEK inhibitor (PD0325901) through the lactating mother's milk prevented NF1-OPG formation [201]. A recent report attributed the predominance of OPG formation in girls to higher levels of glial interleukin-1 β , which can be suppressed by IL-1 β neutralization and leuprolide-mediated estrogen suppression [192,202]. However, it is important to note that the formation of the optic chiasm in mice differs from humans [181] and these tissue-restrictive tumors in mice do not fully replicate the complex pathology of NF1 patients [203]. Additionally, humans have a different proportion of ipsilateral and contralateral projecting RGCs, with ~50% of RGC axons decussating to the contralateral optic tract, while in rodents, only a few RGCs contribute to binocular vision (with ~95–97% of RGCs projecting contralaterally [204-206]).

Large animal models of NF1 offer a better anatomical comparison to understand the NF1 pathology in humans. Genetically engineered porcine models, such as the *Nf1*^{+/*R*1947X} minipigs, share major clinical NF1 features. Electron microscopic evaluation of the optic nerves demonstrated significant demyelination and OPG formation that was confirmed by MRI and CT scans [207,208]. Additionally, their comparable eye and optic nerve size to humans can facilitate the development of new imaging approaches for diagnosis and the testing of novel surgical modalities. Their body size, metabolism and lifespan make them an ideal preclinical model for longitudinal studies, pharmacological tests, and drug dose optimization studies [203]. Large animal models may facilitate translational studies, acting as an intermediate between small rodents and humans [180]. Similarly, cases of spontaneous NF-like manifestations in large animals, such us cattle and dogs [203,209,210], are highly valuable because they can provide insights into the natural malignant transformation of some tumors, contrasting with genetically engineered models [211].

From a different perspective, the zebrafish model can facilitate large-scale experiments for treatment screening, as generating transgenic lines is cost-effective since they are easy to handle and have a high fertility rate [212,213]. The drosophila model can also contribute to large-scale testing; however, these studies typically focus on peripheral nerves, social abilities, and development [214,215]. In addition to animal models, cell culture methods also enable high-throughput screening, which could be particularly valuable when using cells developed from patients to generate compact LGGs [216].

An integrative approach requires collaboration between clinicians and researchers. This collaboration not only enriches our understanding of the disease but also bridges the gap between clinical observations and laboratory advancements. Furthermore, neuroscientists are exploring novel strategies to promote RGC survival and regeneration, including neuroprotective drugs [217], gene therapy [218], or stem cell transplantation [219,220]. These approaches hold promise for improving the vision of NF1 patients; however, most are still in the experimental stage, requiring further research to determine their safety and efficacy.

6. Concluding Remarks

The study and treatment of NF1 pose significant challenges owing to its variable clinical presentation and biological complexity. Clinically, NF1 exhibits remarkable heterogeneity, manifesting diverse symptoms and complications, with variability in penetrance and unpredictable progression of associated OPGs and retinal abnormalities. The major challenge in understanding NF1 pathophysiology arises from the extensive genetic and phenotypic variability, coupled with the absence of clear associations with visual deficits. Further complicating NF1-related retinal research is the lack of well-defined biomarkers that can differentiate between asymptomatic and symptomatic OPGs that lead to vision loss. Unlike systemic aspects of NF1 that are accessible and more easily monitored, the intricate structure and function of the retina and brain demand sophisticated imaging techniques and functional assessment, which may not translate readily into easily measurable biomarkers. Despite these challenges, progress in genetic research, development of new imaging technologies, and collaborative efforts are gradually enhancing our understanding of NF1.

Ethical considerations introduce additional complexity, particularly in dealing with pediatric populations who cannot consent to clinical investigations for themselves. The hereditary nature of NF1 requires careful navigation of informed consent from parents/guardians and privacy concerns. This highlights the need for an ethical framework that respects the rights and well-being of patients, making the assembly of large patient cohorts difficult, and limiting robust data analysis. This emphasizes the importance of collaborative efforts across research centers to consolidate data and share insights.

While animal models have been developed to better understand the mechanism of NF1 pathology and to design new therapeutic approaches, replicating the full spectrum of abnormalities to the retina, optic nerve, and visual pathways observed in humans remains a significant challenge. While mouse lines can mimic certain features of the NF1 pathology, species-specific differences in ocular/brain anatomy and physiology may limit translation to the human condition. Large animal models such as minipigs exhibit a closer resemblance to human anatomy and physiology. Newer models are being developed to better replicate human NF1 pathology, and to shed light on NF1-related visual impairment.

Integration of clinical observations, advanced imaging technologies, and molecular analyses requires collaboration among diverse fields, including ophthalmology, genetics, neurology, and basic science research. Moreover, understanding the interplay between genetic and environmental factors in the development and progression of retinal abnormalities associated with NF1 is crucial. Untangling these interactions is essential to understanding the underlying causes of many of the NF1-related retinal manifestations. Through this integrative review, we summarize the ocular facets of this inherited disorder and encourage synergistic collaboration to discover therapeutic interventions to mitigate visual loss in NF1-affected individuals.

Author Contributions: Conceptualization, K.J.M. and F.M.N.-N.; writing—original draft, F.M.N.-N.; visualization, F.M.N.-N.; writing—review and editing, K.J.M., F.Q., S.F.S. and F.M.N.-N. All authors have read and agreed to the published version of the manuscript.

Funding: This research was supported [in part] by Intramural Research Program of the National Institutes of Health, National Eye Institute.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Conflicts of Interest: The authors declare no conflicts of interest.

References

- 1. Ricker, C.A.; Pan, Y.; Gutmann, D.H.; Keller, C. Challenges in Drug Discovery for Neurofibromatosis Type 1-Associated Low-Grade Glioma. *Front. Oncol.* **2016**, *6*, 259. [CrossRef]
- 2. Riccardi, V.M. Neurofibromatosis: Past, Present, and Future. N. Engl. J. Med. 1991, 324, 1283–1285. [CrossRef]
- 3. Wang, Z.; Liu, Y. Research Update and Recent Developments in the Management of Scoliosis in Neurofibromatosis Type 1. *Orthopedics* **2010**, *33*, 335–341. [CrossRef]
- 4. Ars, E.; Kruyer, H.; Morell, M.; Pros, E.; Serra, E.; Ravella, A.; Estivill, X.; Lázaro, C. Recurrent Mutations in the NF1 Gene Are Common among Neurofibromatosis Type 1 Patients. *J. Med. Genet.* 2003, 40, e82. [CrossRef]
- 5. Cichowski, K.; Jacks, T. NF1 Tumor Suppressor Gene Function: Narrowing the GAP. Cell 2001, 104, 593–604. [CrossRef]

- Yap, Y.-S.; McPherson, J.R.; Ong, C.-K.; Rozen, S.G.; Teh, B.-T.; Lee, A.S.G.; Callen, D.F. The NF1 Gene Revisited—From Bench to Bedside. Oncotarget 2014, 5, 5873–5892. [CrossRef]
- Ratner, N.; Miller, S.J. A RASopathy Gene Commonly Mutated in Cancer: The Neurofibromatosis Type 1 Tumour Suppressor. Nat. Rev. Cancer 2015, 15, 290–301. [CrossRef]
- Listernick, R.; Ferner, R.E.; Liu, G.T.; Gutmann, D.H. Optic Pathway Gliomas in Neurofibromatosis-1: Controversies and Recommendations. *Ann. Neurol.* 2007, 61, 189–198. [CrossRef]
- 9. Nix, J.S.; Blakeley, J.; Rodriguez, F.J. An Update on the Central Nervous System Manifestations of Neurofibromatosis Type 1. *Acta Neuropathol.* **2020**, 139, 625–641. [CrossRef]
- 10. Vitale, M.G.; Guha, A.; Skaggs, D.L. Orthopaedic Manifestations of Neurofibromatosis in Children: An Update. *Clin. Orthop. Relat. Res.* 2002, 401, 107–118. [CrossRef]
- 11. Toro, G.; Santoro, C.; Ambrosio, D.; Landi, G.; Scilipoti, M.; Moretti, A.; Paoletta, M.; Liguori, S.; Schiavone Panni, A.; Picariello, S.; et al. Natural History of Scoliosis in Children with NF1: An Observation Study. *Healthcare* **2021**, *9*, 881. [CrossRef]
- 12. Schindera, C.; Wingeier, K.; Goeggel Simonetti, B.; Diepold, M.; Nauer, C.B.; Fleischhauer, J.; Steinlin, M. Macrocephaly in Neurofibromatosis Type 1: A Sign Post for Optic Pathway Gliomas? *Childs Nerv. Syst.* **2011**, *27*, 2107–2111. [CrossRef]
- 13. Jett, K.; Friedman, J.M. Clinical and Genetic Aspects of Neurofibromatosis 1. Genet. Med. 2010, 12, 1–11. [CrossRef]
- North, K.N.; Riccardi, V.; Samango-Sprouse, C.; Ferner, R.; Moore, B.; Legius, E.; Ratner, N.; Denckla, M.B. Cognitive Function and Academic Performance in Neurofibromatosis. 1: Consensus Statement from the NF1 Cognitive Disorders Task Force. *Neurology* 1997, 48, 1121–1127. [CrossRef]
- 15. Hyman, S.L.; Shores, A.; North, K.N. The Nature and Frequency of Cognitive Deficits in Children with Neurofibromatosis Type 1. *Neurology* **2005**, *65*, 1037–1044. [CrossRef]
- 16. Torres Nupan, M.M.; Velez Van Meerbeke, A.; López Cabra, C.A.; Herrera Gomez, P.M. Cognitive and Behavioral Disorders in Children with Neurofibromatosis Type 1. *Front. Pediatr.* **2017**, *5*, 227. [CrossRef]
- Fisher, M.J.; Avery, R.A.; Allen, J.C.; Ardern-Holmes, S.L.; Bilaniuk, L.T.; Ferner, R.E.; Gutmann, D.H.; Listernick, R.; Martin, S.; Ullrich, N.J.; et al. Functional Outcome Measures for NF1-Associated Optic Pathway Glioma Clinical Trials. *Neurology* 2013, *81*, S15–S24. [CrossRef]
- 18. Stasheff, S.F.; Nadal-Nicolas, F.; Jecrois, E.; Li, W.; Bornhorst, M.; Zhu, Y. Physiologic Dysfunction, Demyelination, and Retinal Ganglion Cell Loss in Mice with Neurofibromatosis and Optic Pathway Gliomas. *Investig. Ophthalmol. Vis. Sci.* **2019**, *60*, 3101.
- 19. Ramón y Cajal, S. *La Rétine Des Vertébrés*; Typ. de Joseph van In & Cie.: Gradignan, France, 1892; pp. 122–255. Available online: https://books.google.com/books?id=kmBHnQEACAAJ (accessed on 11 February 2024).
- 20. Hubel, D.H.; Wiesel, T.N. Brain Mechanisms of Vision. Sci. Am. 1979, 241, 150–162. [CrossRef]
- Güler, A.D.; Ecker, J.L.; Lall, G.S.; Haq, S.; Altimus, C.M.; Liao, H.-W.; Barnard, A.R.; Cahill, H.; Badea, T.C.; Zhao, H.; et al. Melanopsin Cells Are the Principal Conduits for Rod-Cone Input to Non-Image-Forming Vision. *Nature* 2008, 453, 102–105. [CrossRef]
- Lucas, R.J.; Hattar, S.; Takao, M.; Berson, D.M.; Foster, R.G.; Yau, K.-W. Diminished Pupillary Light Reflex at High Irradiances in Melanopsin-Knockout Mice. Science 2003, 299, 245–247. [CrossRef]
- London, A.; Benhar, I.; Schwartz, M. The Retina as a Window to the Brain-from Eye Research to CNS Disorders. *Nat. Rev. Neurol.* 2013, 9, 44–53. [CrossRef]
- 24. MacLaren, R.E. Re-Establishment of Visual Circuitry after Optic Nerve Regeneration. Eye 1999, 13, 277–284. [CrossRef]
- Rojnueangnit, K.; Xie, J.; Gomes, A.; Sharp, A.; Callens, T.; Chen, Y.; Liu, Y.; Cochran, M.; Abbott, M.-A.; Atkin, J.; et al. High Incidence of Noonan Syndrome Features Including Short Stature and Pulmonic Stenosis in Patients Carrying NF1 Missense Mutations Affecting p.Arg1809: Genotype-Phenotype Correlation. *Hum. Mutat.* 2015, *36*, 1052–1063. [CrossRef]
- Pasmant, E.; Sabbagh, A.; Spurlock, G.; Laurendeau, I.; Grillo, E.; Hamel, M.-J.; Martin, L.; Barbarot, S.; Leheup, B.; Rodriguez, D.; et al. NF1 Microdeletions in Neurofibromatosis Type 1: From Genotype to Phenotype. *Hum. Mutat.* 2010, 31, E1506–E1518. [CrossRef]
- Wang, Q.; Montmain, G.; Ruano, E.; Upadhyaya, M.; Dudley, S.; Liskay, R.M.; Thibodeau, S.N.; Puisieux, A. Neurofibromatosis Type 1 Gene as a Mutational Target in a Mismatch Repair-Deficient Cell Type. *Hum. Genet.* 2003, *112*, 117–123. [CrossRef]
- Kehrer-Sawatzki, H.; Mautner, V.-F.; Cooper, D.N. Emerging Genotype-Phenotype Relationships in Patients with Large NF1 Deletions. *Hum. Genet.* 2017, 136, 349–376. [CrossRef]
- 29. Easton, D.F.; Ponder, M.A.; Huson, S.M.; Ponder, B.A. An Analysis of Variation in Expression of Neurofibromatosis (NF) Type 1 (NF1): Evidence for Modifying Genes. *Am. J. Hum. Genet.* **1993**, *53*, 305–313.
- Makino, S.; Tampo, H. Optical Coherence Tomography Imaging of Choroidal Abnormalities in Neurofibromatosis Type 1. *Case Rep. Ophthalmol. Med.* 2013, 2013, 292981. [CrossRef]
- Vagge, A.; Camicione, P.; Pellegrini, M.; Gatti, G.; Capris, P.; Severino, M.; Di Maita, M.; Panarello, S.; Traverso, C.E. Role of Visual Evoked Potentials and Optical Coherence Tomography in the Screening for Optic Pathway Gliomas in Patients with Neurofibromatosis Type I. *Eur. J. Ophthalmol.* 2021, *31*, 698–703. [CrossRef]
- 32. Wang, M.X.; Dillman, J.R.; Guccione, J.; Habiba, A.; Maher, M.; Kamel, S.; Panse, P.M.; Jensen, C.T.; Elsayes, K.M. Neurofibromatosis from Head to Toe: What the Radiologist Needs to Know. *Radiographics* **2022**, *42*, 1123–1144. [CrossRef]
- 33. Huang, M.; Patel, J.; Patel, B.C. Optic Nerve Glioma; StatPearls Publishing: Treasure Island, FL, USA, 2024.

- Eid, H.; Crevier-Sorbo, G.; Aldraihem, A.; Menegotto, F.; Wilson, N. Neurofibromatosis Type 1: Description of a Novel Diagnostic Scoring System in Pediatric Optic Nerve Glioma. *AJR Am. J. Roentgenol.* 2019, 212, 892–898. [CrossRef]
- Beres, S.J.; Avery, R.A. Optic Pathway Gliomas Secondary to Neurofibromatosis Type 1. Semin Pediatr. Neurol. 2017, 24, 92–99. [CrossRef]
- Zeid, J.L.; Charrow, J.; Sandu, M.; Goldman, S.; Listernick, R. Orbital Optic Nerve Gliomas in Children with Neurofibromatosis Type 1. J. AAPOS 2006, 10, 534–539. [CrossRef]
- Pisapia, J.M.; Akbari, H.; Rozycki, M.; Thawani, J.P.; Storm, P.B.; Avery, R.A.; Vossough, A.; Fisher, M.J.; Heuer, G.G.; Davatzikos, C. Predicting Pediatric Optic Pathway Glioma Progression Using Advanced Magnetic Resonance Image Analysis and Machine Learning. *Neuro-Oncol. Adv.* 2020, 2, vdaa090. [CrossRef]
- Cummings, T.J.; Provenzale, J.M.; Hunter, S.B.; Friedman, A.H.; Klintworth, G.K.; Bigner, S.H.; McLendon, R.E. Gliomas of the Optic Nerve: Histological, Immunohistochemical (MIB-1 and P53), and MRI Analysis. *Acta Neuropathol.* 2000, *99*, 563–570. [CrossRef]
- 39. Perry, A.; Wesseling, P. Histologic Classification of Gliomas. Handb. Clin. Neurol. 2016, 134, 71–95. [CrossRef]
- Nadal-Nicolas, F.M.; Jimenez-Lopez, M.; Sobrado-Calvo, P.; Nieto-Lopez, L.; Canovas-Martinez, I.; Salinas-Navarro, M.; Vidal-Sanz, M.; Agudo, M. Brn3a as a Marker of Retinal Ganglion Cells: Qualitative and Quantitative Time Course Studies in Naive and Optic Nerve-Injured Retinas. *Investig. Ophthalmol. Vis. Sci.* 2009, 50, 3860–3868. [CrossRef]
- 41. Nadal-Nicolás, F.M.; Galindo-Romero, C.; Lucas-Ruiz, F.; Marsh-Amstrong, N.; Li, W.; Vidal-Sanz, M.; Agudo-Barriuso, M. Pan-Retinal Ganglion Cell Markers in Mice, Rats, and Rhesus Macaques. *Zool. Res.* **2023**, *44*, 226–248. [CrossRef]
- 42. Gutmann, D.H.; Ferner, R.E.; Listernick, R.H.; Korf, B.R.; Wolters, P.L.; Johnson, K.J. Neurofibromatosis Type 1. *Nat. Rev. Dis. Primers* 2017, 3, 17004. [CrossRef]
- 43. Huson, S.; Jones, D.; Beck, L. Ophthalmic Manifestations of Neurofibromatosis. Br. J. Ophthalmol. 1987, 71, 235–238. [CrossRef]
- Maharaj, A.; Singh, V.R.; Lalchan, S.A. Lisch and the Importance of His Nodules. *West Indian Med. J.* 2014, 63, 799–802. [CrossRef]
 Hernáiz Driever, P.; von Hornstein, S.; Pietsch, T.; Kortmann, R.; Warmuth-Metz, M.; Emser, A.; Gnekow, A.K. Natural History and Management of Low-Grade Glioma in NF-1 Children. *J. Neuro-Oncol.* 2010, 100, 199–207. [CrossRef]
- Fisher, M.J.; Jones, D.T.W.; Li, Y.; Guo, X.; Sonawane, P.S.; Waanders, A.J.; Phillips, J.J.; Weiss, W.A.; Resnick, A.C.; Gosline, S.; et al. Integrated Molecular and Clinical Analysis of Low-Grade Gliomas in Children with Neurofibromatosis Type 1 (NF1). *Acta Neuropathol.* 2021, 141, 605–617. [CrossRef]
- Lohkamp, L.-N.; Parkin, P.; Puran, A.; Bartels, U.K.; Bouffet, E.; Tabori, U.; Rutka, J.T. Optic Pathway Glioma in Children with Neurofibromatosis Type 1: A Multidisciplinary Entity, Posing Dilemmas in Diagnosis and Management Multidisciplinary Management of Optic Pathway Glioma in Children with Neurofibromatosis Type 1. *Front. Surg.* 2022, *9*, 886697. [CrossRef]
- 48. Shofty, B.; Ben Sira, L.; Constantini, S. Neurofibromatosis 1-Associated Optic Pathway Gliomas. *Childs Nerv. Syst.* 2020, *36*, 2351–2361. [CrossRef]
- Taylor, T.; Jaspan, T.; Milano, G.; Gregson, R.; Parker, T.; Ritzmann, T.; Benson, C.; Walker, D. PLAN Study Group Radiological Classification of Optic Pathway Gliomas: Experience of a Modified Functional Classification System. *Br. J. Radiol.* 2008, *81*, 761–766. [CrossRef]
- 50. Listernick, R.; Charrow, J.; Greenwald, M.; Mets, M. Natural History of Optic Pathway Tumors in Children with Neurofibromatosis Type 1: A Longitudinal Study. J. Pediatr. **1994**, 125, 63–66. [CrossRef]
- 51. Rodriguez, F.J.; Perry, A.; Gutmann, D.H.; O'Neill, B.P.; Leonard, J.; Bryant, S.; Giannini, C. Gliomas in Neurofibromatosis Type 1: A Clinicopathologic Study of 100 Patients. *J. Neuropathol. Exp. Neurol.* **2008**, *67*, 240–249. [CrossRef]
- Azizi, A.A.; Walker, D.A.; Liu, J.-F.; Sehested, A.; Jaspan, T.; Pemp, B.; Simmons, I.; Ferner, R.; Grill, J.; Hargrave, D.; et al. NF1 Optic Pathway Glioma: Analyzing Risk Factors for Visual Outcome and Indications to Treat. *Neuro-Oncology* 2021, 23, 100–111. [CrossRef]
- 53. Angelova-Toshkina, D.; Decker, J.A.; Traunwieser, T.; Holzapfel, J.; Bette, S.; Huber, S.; Schimmel, M.; Vollert, K.; Bison, B.; Kröncke, T.; et al. Comprehensive Neurological Evaluation of a Cohort of Patients with Neurofibromatosis Type 1 from a Single Institution. *Eur. J. Paediatr. Neurol.* **2023**, *43*, 52–61. [CrossRef]
- 54. Parsa, C.F.; Hoyt, C.S.; Lesser, R.L.; Weinstein, J.M.; Strother, C.M.; Muci-Mendoza, R.; Ramella, M.; Manor, R.S.; Fletcher, W.A.; Repka, M.X.; et al. Spontaneous Regression of Optic Gliomas: Thirteen Cases Documented by Serial Neuroimaging. *Arch. Oph-thalmol.* **2001**, *119*, 516–529. [CrossRef]
- 55. Senthilkumar, V.A.; Tripathy, K. Lisch Nodules; StatPearls Publishing: Treasure Island, FL, USA, 2024.
- 56. Papageorgiou, E.; Tsironi-Malizou, E. Types of Homonymous Visual Field Defects. In *Homonymous Visual Field Defects*; Skorkovská, K., Ed.; Springer: Cham, Switzerland, 2017; pp. 65–94. ISBN 978-3-319-52282-1.
- 57. Walker, D.A.; Aquilina, K.; Spoudeas, H.; Pilotto, C.; Gan, H.-W.; Meijer, L. A New Era for Optic Pathway Glioma: A Developmental Brain Tumor with Life-Long Health Consequences. *Front. Pediatr.* **2023**, *11*, 1038937. [CrossRef]
- 58. Silva, M.M.; Goldman, S.; Keating, G.; Marymont, M.A.; Kalapurakal, J.; Tomita, T. Optic Pathway Hypothalamic Gliomas in Children under Three Years of Age: The Role of Chemotherapy. *Pediatr. Neurosurg.* **2000**, *33*, 151–158. [CrossRef]
- Fuss, M.; Hug, E.B.; Schaefer, R.A.; Nevinny-Stickel, M.; Miller, D.W.; Slater, J.M.; Slater, J.D. Proton Radiation Therapy (PRT) for Pediatric Optic Pathway Gliomas: Comparison with 3D Planned Conventional Photons and a Standard Photon Technique. *Int. J. Radiat. Oncol. Biol. Phys.* 1999, 45, 1117–1126. [CrossRef]

- Balcer, L.J.; Liu, G.T.; Heller, G.; Bilaniuk, L.; Volpe, N.J.; Galetta, S.L.; Molloy, P.T.; Phillips, P.C.; Janss, A.J.; Vaughn, S.; et al. Visual Loss in Children with Neurofibromatosis Type 1 and Optic Pathway Gliomas: Relation to Tumor Location by Magnetic Resonance Imaging. *Am. J. Ophthalmol.* 2001, 131, 442–445. [CrossRef]
- Falzon, K.; Drimtzias, E.; Picton, S.; Simmons, I. Visual Outcomes after Chemotherapy for Optic Pathway Glioma in Children with and without Neurofibromatosis Type 1: Results of the International Society of Paediatric Oncology (SIOP) Low-Grade Glioma 2004 Trial UK Cohort. Br. J. Ophthalmol. 2018, 102, 1367–1371. [CrossRef]
- 62. Lobbous, M.; Bernstock, J.D.; Coffee, E.; Friedman, G.K.; Metrock, L.K.; Chagoya, G.; Elsayed, G.; Nakano, I.; Hackney, J.R.; Korf, B.R.; et al. An Update on Neurofibromatosis Type 1-Associated Gliomas. *Cancers* **2020**, *12*, 114. [CrossRef]
- 63. Friedman, J.M. Neurofibromatosis 1. In *GeneReviews*[®]; Adam, M.P., Feldman, J., Mirzaa, G.M., Pagon, R.A., Wallace, S.E., Bean, L.J., Gripp, K.W., Amemiya, A., Eds.; University of Washington: Seattle, WA, USA, 1993.
- Parrozzani, R.; Pilotto, E.; Clementi, M.; Frizziero, L.; Leonardi, F.; Convento, E.; Miglionico, G.; Pulze, S.; Perrini, P.; Trevisson, E.; et al. Retinal Vascular Abnormalities in a Large Cohort of Patients Affected by Neurofibromatosis Type 1: A Study Using Optical Coherence Tomography Angiography. *Retina* 2018, *38*, 585–593. [CrossRef]
- 65. Moramarco, A.; Miraglia, E.; Mallone, F.; Roberti, V.; Iacovino, C.; Bruscolini, A.; Giustolisi, R.; Giustini, S. Retinal Microvascular Abnormalities in Neurofibromatosis Type 1. *Br. J. Ophthalmol.* **2019**, *103*, 1590–1594. [CrossRef]
- 66. Mallone, F.; Lucchino, L.; Giustini, S.; Lambiase, A.; Moramarco, A. An Update on Choroidal Abnormalities and Retinal Microvascular Changes in Neurofibromatosis Type 1. *Orphanet J. Rare Dis.* **2022**, *17*, 223. [CrossRef]
- 67. Hoyt, W.F.; Luis, O. The Primate Chiasm. Details of Visual Fiber Organization Studied by Silver Impregnation Techniques. *Arch. Ophthalmol.* **1963**, *70*, 69–85. [CrossRef]
- Rodriguez, F.J.; Ligon, A.H.; Horkayne-Szakaly, I.; Rushing, E.J.; Ligon, K.L.; Vena, N.; Garcia, D.I.; Cameron, J.D.; Eberhart, C.G. BRAF Duplications and MAPK Pathway Activation Are Frequent in Gliomas of the Optic Nerve Proper. *J. Neuropathol. Exp. Neurol.* 2012, *71*, 789–794. [CrossRef]
- 69. Robert-Boire, V.; Rosca, L.; Samson, Y.; Ospina, L.H.; Perreault, S. Clinical Presentation and Outcome of Patients with Optic Pathway Glioma. *Pediatr. Neurol.* 2017, 75, 55–60. [CrossRef]
- Modrzejewska, M.; Olejnik-Wojciechowska, J.; Roszyk, A.; Szychot, E.; Konczak, T.D.; Szemitko, M.; Peregud-Pogorzelski, J.W. Optic Pathway Gliomas in Pediatric Population-Current Approach in Diagnosis and Management: Literature Review. J. Clin. Med. 2023, 12, 6709. [CrossRef]
- 71. Ronsley, R.; Hounjet, C.D.; Cheng, S.; Rassekh, S.R.; Duncan, W.J.; Dunham, C.; Gardiner, J.; Ghag, A.; Ludemann, J.P.; Wensley, D.; et al. Trametinib Therapy for Children with Neurofibromatosis Type 1 and Life-Threatening Plexiform Neurofibroma or Treatment-Refractory Low-Grade Glioma. *Cancer Med.* 2021, 10, 3556–3564. [CrossRef]
- Manoharan, N.; Choi, J.; Chordas, C.; Zimmerman, M.A.; Scully, J.; Clymer, J.; Filbin, M.; Ullrich, N.J.; Bandopadhayay, P.; Chi, S.N.; et al. Trametinib for the Treatment of Recurrent/Progressive Pediatric Low-Grade Glioma. *J. Neuro-Oncol.* 2020, 149, 253–262. [CrossRef]
- Jones, D.T.W.; Hutter, B.; Jäger, N.; Korshunov, A.; Kool, M.; Warnatz, H.-J.; Zichner, T.; Lambert, S.R.; Ryzhova, M.; Quang, D.A.K.; et al. Recurrent Somatic Alterations of FGFR1 and NTRK2 in Pilocytic Astrocytoma. *Nat. Genet.* 2013, 45, 927–932. [CrossRef]
- 74. Sharma, M.K.; Zehnbauer, B.A.; Watson, M.A.; Gutmann, D.H. RAS Pathway Activation and an Oncogenic RAS Mutation in Sporadic Pilocytic Astrocytoma. *Neurology* **2005**, *65*, 1335–1336. [CrossRef]
- Czyzyk, E.; Jóźwiak, S.; Roszkowski, M.; Schwartz, R.A. Optic Pathway Gliomas in Children with and without Neurofibromatosis 1. J. Child Neurol. 2003, 18, 471–478. [CrossRef]
- 76. Listernick, R.; Darling, C.; Greenwald, M.; Strauss, L.; Charrow, J. Optic Pathway Tumors in Children: The Effect of Neurofibromatosis Type 1 on Clinical Manifestations and Natural History. *J. Pediatr.* **1995**, 127, 718–722. [CrossRef]
- 77. Rasool, N.; Odel, J.G.; Kazim, M. Optic Pathway Glioma of Childhood. Curr. Opin. Ophthalmol. 2017, 28, 289-295. [CrossRef]
- 78. Singhal, S.; Birch, J.M.; Kerr, B.; Lashford, L.; Evans, D.G.R. Neurofibromatosis Type 1 and Sporadic Optic Gliomas. *Arch. Dis. Child* 2002, *87*, 65–70. [CrossRef]
- 79. Astrup, J. Natural History and Clinical Management of Optic Pathway Glioma. Br. J. Neurosurg. 2003, 17, 327–335. [CrossRef]
- 80. Chateil, J.F.; Soussotte, C.; Pédespan, J.M.; Brun, M.; Le Manh, C.; Diard, F. MRI and Clinical Differences between Optic Pathway Tumours in Children with and without Neurofibromatosis. *Br. J. Radiol.* **2001**, *74*, 24–31. [CrossRef]
- 81. Shamji, M.F.; Benoit, B.G. Syndromic and Sporadic Pediatric Optic Pathway Gliomas: Review of Clinical and Histopathological Differences and Treatment Implications. *Neurosurg. Focus* **2007**, *23*, E3. [CrossRef]
- Kornreich, L.; Blaser, S.; Schwarz, M.; Shuper, A.; Vishne, T.H.; Cohen, I.J.; Faingold, R.; Michovitz, S.; Koplewitz, B.; Horev, G. Optic Pathway Glioma: Correlation of Imaging Findings with the Presence of Neurofibromatosis. *AJNR Am. J. Neuroradiol.* 2001, 22, 1963–1969.
- Grill, J.; Laithier, V.; Rodriguez, D.; Raquin, M.A.; Pierre-Kahn, A.; Kalifa, C. When Do Children with Optic Pathway Tumours Need Treatment? An Oncological Perspective in 106 Patients Treated in a Single Centre. *Eur. J. Pediatr.* 2000, 159, 692–696. [CrossRef]
- 84. Wan, M.J.; Ullrich, N.J.; Manley, P.E.; Kieran, M.W.; Goumnerova, L.C.; Heidary, G. Long-Term Visual Outcomes of Optic Pathway Gliomas in Pediatric Patients without Neurofibromatosis Type 1. *J. Neuro-Oncol.* **2016**, *129*, 173–178. [CrossRef]

- 85. NIH. National Institutes of Health Consensus Development Conference Statement: Neurofibromatosis, Bethesda, Md., USA, July 13–15, 1987. *Neurofibromatosis* **1988**, *1*, 172–178.
- Kang, E.; Yoon, H.M.; Lee, B.H. Neurofibromatosis Type I: Points to Be Considered by General Pediatricians. *Clin. Exp. Pediatr.* 2021, 64, 149–156. [CrossRef]
- de Blank, P.M.K.; Fisher, M.J.; Liu, G.T.; Gutmann, D.H.; Listernick, R.; Ferner, R.E.; Avery, R.A. Optic Pathway Gliomas in Neurofibromatosis Type 1: An Update: Surveillance, Treatment Indications, and Biomarkers of Vision. *J. Neuroophthalmol.* 2017, 37 (Suppl. S1), S23–S32. [CrossRef]
- Tang, Y.; Gutmann, D.H. Neurofibromatosis Type 1-Associated Optic Pathway Gliomas: Current Challenges and Future Prospects. *Cancer Manag. Res.* 2023, 15, 667–681. [CrossRef]
- Lubs, M.L.; Bauer, M.S.; Formas, M.E.; Djokic, B. Lisch Nodules in Neurofibromatosis Type 1. N. Engl. J. Med. 1991, 324, 1264–1266.
 [CrossRef]
- 90. Daoudi, C.; Daoudi, R. Lisch nodules in Von Recklinghausen disease. Pan Afr. Med. J. 2014, 19, 173. [CrossRef]
- Binning, M.J.; Liu, J.K.; Kestle, J.R.W.; Brockmeyer, D.L.; Walker, M.L. Optic Pathway Gliomas: A Review. Neurosurg. Focus 2007, 23, E2. [CrossRef]
- 92. Fletcher, W.A.; Imes, R.K.; Hoyt, W.F. Chiasmal Gliomas: Appearance and Long-Term Changes Demonstrated by Computerized Tomography. *J. Neurosurg.* **1986**, *65*, 154–159. [CrossRef]
- 93. Moreno, L.; Bautista, F.; Ashley, S.; Duncan, C.; Zacharoulis, S. Does Chemotherapy Affect the Visual Outcome in Children with Optic Pathway Glioma? A Systematic Review of the Evidence. *Eur. J. Cancer* **2010**, *46*, 2253–2259. [CrossRef]
- Maloney, E.; Perez, F.A.; Iyer, R.S.; Otto, R.K.; Wright, J.N.; Menashe, S.J.; Hippe, D.S.; Shaw, D.W.W.; Stanescu, A.L. Non-Inferiority of a Non-Gadolinium-Enhanced Magnetic Resonance Imaging Follow-up Protocol for Isolated Optic Pathway Gliomas. *Pediatr. Radiol.* 2022, 52, 539–548. [CrossRef]
- 95. Walrath, J.D.; Engelbert, M.; Kazim, M. Magnetic Resonance Imaging Evidence of Optic Nerve Glioma Progression into and beyond the Optic Chiasm. *Ophthalmic Plast. Reconstr. Surg.* **2008**, *24*, 473–475. [CrossRef]
- 96. Fisher, M.J.; Loguidice, M.; Gutmann, D.H.; Listernick, R.; Ferner, R.E.; Ullrich, N.J.; Packer, R.J.; Tabori, U.; Hoffman, R.O.; Ardern-Holmes, S.L.; et al. Visual Outcomes in Children with Neurofibromatosis Type 1-Associated Optic Pathway Glioma Following Chemotherapy: A Multicenter Retrospective Analysis. *Neuro-Oncol.* 2012, 14, 790–797. [CrossRef] [PubMed]
- Wei, C.-J.; Yan, C.; Tang, Y.; Wang, W.; Gu, Y.-H.; Ren, J.-Y.; Cui, X.-W.; Lian, X.; Liu, J.; Wang, H.-J.; et al. Computed Tomography-Based Differentiation of Benign and Malignant Craniofacial Lesions in Neurofibromatosis Type I Patients: A Machine Learning Approach. *Front. Oncol.* 2020, 10, 1192. [CrossRef] [PubMed]
- 98. Avery, R.A.; Mansoor, A.; Idrees, R.; Trimboli-Heidler, C.; Ishikawa, H.; Packer, R.J.; Linguraru, M.G. Optic Pathway Glioma Volume Predicts Retinal Axon Degeneration in Neurofibromatosis Type 1. *Neurology* **2016**, *87*, 2403–2407. [CrossRef] [PubMed]
- Blanchard, G.; Lafforgue, M.-P.; Lion-François, L.; Kemlin, I.; Rodriguez, D.; Castelnau, P.; Carneiro, M.; Meyer, P.; Rivier, F.; Barbarot, S.; et al. Systematic MRI in NF1 Children under Six Years of Age for the Diagnosis of Optic Pathway Gliomas. Study and Outcome of a French Cohort. *Eur. J. Paediatr. Neurol.* 2016, 20, 275–281. [CrossRef] [PubMed]
- 100. Cassina, M.; Frizziero, L.; Opocher, E.; Parrozzani, R.; Sorrentino, U.; Viscardi, E.; Miglionico, G.; Midena, E.; Clementi, M.; Trevisson, E. Optic Pathway Glioma in Type 1 Neurofibromatosis: Review of Its Pathogenesis, Diagnostic Assessment, and Treatment Recommendations. *Cancers* 2019, *11*, 1790. [CrossRef] [PubMed]
- 101. Parrozzani, R.; Clementi, M.; Kotsafti, O.; Miglionico, G.; Trevisson, E.; Orlando, G.; Pilotto, E.; Midena, E. Optical Coherence Tomography in the Diagnosis of Optic Pathway Gliomas. *Investig. Ophthalmol. Vis. Sci.* **2013**, *54*, 8112–8118. [CrossRef] [PubMed]
- 102. Avery, R.A.; Cnaan, A.; Schuman, J.S.; Trimboli-Heidler, C.; Chen, C.-L.; Packer, R.J.; Ishikawa, H. Longitudinal Change of Circumpapillary Retinal Nerve Fiber Layer Thickness in Children with Optic Pathway Gliomas. *Am. J. Ophthalmol.* 2015, 160, 944–952.e1. [CrossRef] [PubMed]
- 103. Avery, R.A.; Liu, G.T.; Fisher, M.J.; Quinn, G.E.; Belasco, J.B.; Phillips, P.C.; Maguire, M.G.; Balcer, L.J. Retinal Nerve Fiber Layer Thickness in Children with Optic Pathway Gliomas. Am. J. Ophthalmol. 2011, 151, 542–549.e2. [CrossRef] [PubMed]
- 104. Xiao, X.; Zhao, T.; Miyagishima, K.J.; Chen, S.; Li, W.; Nadal-Nicolás, F.M. Establishing the Ground Squirrel as a Superb Model for Retinal Ganglion Cell Disorders and Optic Neuropathies. *Lab. Investig.* 2021, 101, 1289–1303. [CrossRef]
- 105. Nadal-Nicolás, F.M.; Miyagishima, K.J.; Li, W. In Search for the "Idyllic" Animal Model to Evaluate Ocular Pathologies and Translate New Therapies to Improve Human Health. *Neural Regen. Res.* **2022**, *17*, 2697–2699. [CrossRef]
- 106. Barkana, Y.; Burgansky-Eliash, Z.; Gerber, Y.; Melamed, S.; Neudorfer, M.; Avni, I.; Bartov, E.; Morad, Y. Inter-Device Variability of the Stratus Optical Coherence Tomography. *Am. J. Ophthalmol.* **2009**, *147*, 260–266. [CrossRef]
- 107. Yang, H.; Lee, H.S.; Bae, H.W.; Seong, G.J.; Kim, C.Y.; Lee, S.Y. Effect of Image Quality Fluctuations on the Repeatability of Thickness Measurements in Swept-Source Optical Coherence Tomography. *Sci. Rep.* 2020, 10, 13897. [CrossRef]
- 108. Rovere, G.; Nadal-Nicolás, F.M.; Agudo-Barriuso, M.; Sobrado-Calvo, P.; Nieto-López, L.; Nucci, C.; Villegas-Pérez, M.P.; Vidal-Sanz, M. Comparison of Retinal Nerve Fiber Layer Thinning and Retinal Ganglion Cell Loss after Optic Nerve Transection in Adult Albino Rats. *Investig. Ophthalmol. Vis. Sci.* 2015, *56*, 4487–4498. [CrossRef]
- Moramarco, A.; Giustini, S.; Nofroni, I.; Mallone, F.; Miraglia, E.; Iacovino, C.; Calvieri, S.; Lambiase, A. Near-Infrared Imaging: An in Vivo, Non-Invasive Diagnostic Tool in Neurofibromatosis Type 1. *Graefes Arch. Clin. Exp. Ophthalmol.* 2018, 256, 307–311. [CrossRef]

- 110. Ratnam, K.; Carroll, J.; Porco, T.C.; Duncan, J.L.; Roorda, A. Relationship between Foveal Cone Structure and Clinical Measures of Visual Function in Patients with Inherited Retinal Degenerations. *Investig. Ophthalmol. Vis. Sci.* 2013, *54*, 5836–5847. [CrossRef]
- 111. Foote, K.G.; Loumou, P.; Griffin, S.; Qin, J.; Ratnam, K.; Porco, T.C.; Roorda, A.; Duncan, J.L. Relationship Between Foveal Cone Structure and Visual Acuity Measured with Adaptive Optics Scanning Laser Ophthalmoscopy in Retinal Degeneration. *Investig. Ophthalmol. Vis. Sci.* 2018, 59, 3385–3393. [CrossRef]
- 112. Bensinger, E.; Rinella, N.; Saud, A.; Loumou, P.; Ratnam, K.; Griffin, S.; Qin, J.; Porco, T.C.; Roorda, A.; Duncan, J.L. Loss of Foveal Cone Structure Precedes Loss of Visual Acuity in Patients with Rod-Cone Degeneration. *Investig. Ophthalmol. Vis. Sci.* 2019, 60, 3187–3196. [CrossRef]
- 113. North, K.; Cochineas, C.; Tang, E.; Fagan, E. Optic Gliomas in Neurofibromatosis Type 1: Role of Visual Evoked Potentials. *Pediatr. Neurol.* **1994**, *10*, 117–123. [CrossRef]
- 114. Falsini, B.; Ziccardi, L.; Lazzareschi, I.; Ruggiero, A.; Placentino, L.; Dickmann, A.; Liotti, L.; Piccardi, M.; Balestrazzi, E.; Colosimo, C.; et al. Longitudinal Assessment of Childhood Optic Gliomas: Relationship between Flicker Visual Evoked Potentials and Magnetic Resonance Imaging Findings. J. Neurooncol. 2008, 88, 87–96. [CrossRef]
- Kelly, J.P.; Weiss, A.H. Detection of Tumor Progression in Optic Pathway Glioma with and without Neurofibromatosis Type 1. Neuro-Oncology 2013, 15, 1560–1567. [CrossRef]
- 116. Bowman, R.; Walters, B.; Smith, V.; Prise, K.L.; Handley, S.E.; Green, K.; Mankad, K.; O'Hare, P.; Dahl, C.; Jorgensen, M.; et al. Visual Outcomes and Predictors in Optic Pathway Glioma: A Single Centre Study. *Eye* **2023**, *37*, 1178–1183. [CrossRef]
- 117. Barrea, C.; Vaessen, S.; Bulk, S.; Harvengt, J.; Misson, J.-P. Phenotype-Genotype Correlation in Children with Neurofibromatosis Type 1. *Neuropediatrics* **2018**, *49*, 180–184. [CrossRef]
- 118. Riccardi, V.M. Pathophysiology of Neurofibromatosis. IV. Dermatologic Insights into Heterogeneity and Pathogenesis. *J. Am. Acad. Dermatol.* **1980**, *3*, 157–166. [CrossRef]
- 119. Barker, D.; Wright, E.; Nguyen, K.; Cannon, L.; Fain, P.; Goldgar, D.; Bishop, D.T.; Carey, J.; Baty, B.; Kivlin, J. Gene for von Recklinghausen Neurofibromatosis Is in the Pericentromeric Region of Chromosome 17. *Science* **1987**, *236*, 1100–1102. [CrossRef]
- 120. Koczkowska, M.; Callens, T.; Gomes, A.; Sharp, A.; Chen, Y.; Hicks, A.D.; Aylsworth, A.S.; Azizi, A.A.; Basel, D.G.; Bellus, G.; et al. Expanding the Clinical Phenotype of Individuals with a 3-Bp in-Frame Deletion of the NF1 Gene (c.2970_2972del): An Update of Genotype-Phenotype Correlation. *Genet. Med.* 2019, 21, 867–876. [CrossRef]
- 121. Koczkowska, M.; Chen, Y.; Callens, T.; Gomes, A.; Sharp, A.; Johnson, S.; Hsiao, M.-C.; Chen, Z.; Balasubramanian, M.; Barnett, C.P.; et al. Genotype-Phenotype Correlation in NF1: Evidence for a More Severe Phenotype Associated with Missense Mutations Affecting NF1 Codons 844-848. Am. J. Hum. Genet. 2018, 102, 69–87. [CrossRef]
- 122. Zhu, B.; Zheng, T.; Wang, W.; Gu, Y.; Wei, C.; Li, Q.; Wang, Z. Genotype-Phenotype Correlations of Neurofibromatosis Type 1: A Cross-Sectional Study from a Large Chinese Cohort. J. Neurol. 2023, 271, 1893–1900. [CrossRef]
- 123. Bildirici, Y.; Kocaaga, A.; Karademir-Arslan, C.N.; Yimenicioglu, S. Evaluation of Molecular and Clinical Findings in Children with Neurofibromatosis Type 1: Identification of 15 Novel Variants. *Pediatr. Neurol.* **2023**, *149*, 69–74. [CrossRef]
- 124. Pacot, L.; Sabbagh, A.; Sohier, P.; Hadjadj, D.; Ye, M.; Boland-Auge, A.; Bacq-Daian, D.; Laurendeau, I.; Briand-Suleau, A.; Deleuze, J.-F.; et al. Identification of Potential Common Genetic Modifiers of Neurofibromas: A Genome-Wide Association Study in 1333 Patients with Neurofibromatosis Type 1. *Br. J. Dermatol.* **2024**, *190*, 226–243. [CrossRef]
- 125. Bettegowda, C.; Upadhayaya, M.; Evans, D.G.; Kim, A.; Mathios, D.; Hanemann, C.O. REiNS International Collaboration Genotype-Phenotype Correlations in Neurofibromatosis and Their Potential Clinical Use. *Neurology* **2021**, *97*, S91–S98. [CrossRef]
- 126. Trevisson, E.; Morbidoni, V.; Forzan, M.; Daolio, C.; Fumini, V.; Parrozzani, R.; Cassina, M.; Midena, E.; Salviati, L.; Clementi, M. The Arg1038Gly Missense Variant in the NF1 Gene Causes a Mild Phenotype without Neurofibromas. *Mol. Genet. Genom. Med.* 2019, 7, e616. [CrossRef]
- 127. Kehrer-Sawatzki, H.; Cooper, D.N. Classification of NF1 Microdeletions and Its Importance for Establishing Genotype/Phenotype Correlations in Patients with NF1 Microdeletions. *Hum. Genet.* **2021**, *140*, 1635–1649. [CrossRef]
- Well, L.; Döbel, K.; Kluwe, L.; Bannas, P.; Farschtschi, S.; Adam, G.; Mautner, V.-F.; Salamon, J. Genotype-Phenotype Correlation in Neurofibromatosis Type-1: NF1 Whole Gene Deletions Lead to High Tumor-Burden and Increased Tumor-Growth. *PLoS Genet.* 2021, 17, e1009517. [CrossRef]
- Svensson, C.K.; Drobitch, R.K.; Kloss, K.A. Effect of Glutathione Depletion on the in Vivo Inhibition of Drug Metabolism by Agents Forming an Inactive Cytochrome P-450 Fe(II):Metabolite Complex. Studies with Amiodarone and Troleandomycin. J. Pharm. Sci. 1991, 80, 225–228. [CrossRef]
- 130. Chong, A.L.; Pole, J.D.; Scheinemann, K.; Hukin, J.; Tabori, U.; Huang, A.; Bouffet, E.; Bartels, U. Optic Pathway Gliomas in Adolescence--Time to Challenge Treatment Choices? *Neuro-Oncology* **2013**, *15*, 391–400. [CrossRef]
- Peng, F.; Juhasz, C.; Bhambhani, K.; Wu, D.; Chugani, D.C.; Chugani, H.T. Assessment of Progression and Treatment Response of Optic Pathway Glioma with Positron Emission Tomography Using Alpha-[(11)C]Methyl-L-Tryptophan. *Mol. Imaging Biol.* 2007, 9, 106–109. [CrossRef]
- 132. Revere, K.E.; Katowitz, W.R.; Katowitz, J.A.; Rorke-Adams, L.; Fisher, M.J.; Liu, G.T. Childhood Optic Nerve Glioma: Vision Loss Due to Biopsy. *Ophthalmic Plast. Reconstr. Surg.* 2017, 33, S107–S109. [CrossRef]
- Oderich, G.S.; Sullivan, T.M.; Bower, T.C.; Gloviczki, P.; Miller, D.V.; Babovic-Vuksanovic, D.; Macedo, T.A.; Stanson, A. Vascular Abnormalities in Patients with Neurofibromatosis Syndrome Type I: Clinical Spectrum, Management, and Results. *J. Vasc. Surg.* 2007, 46, 475–484. [CrossRef]

- 134. Hivelin, M.; Plaud, B.; Hemery, F.; Boulat, C.; Ortonne, N.; Valleyrie-Allanore, L.; Wolkenstein, P.; Lantieri, L. Low Rates of Blood Transfusion in Elective Resections of Neurofibromas in a Cohort Study: Neurofibroma Length as a Predictor of Transfusion Requirement. *Plast. Reconstr. Surg.* 2016, 137, 700e–711e. [CrossRef]
- Wang, D.; Ge, L.; Guo, Z.; Li, Y.; Zhu, B.; Wang, W.; Wei, C.; Li, Q.; Wang, Z. Efficacy and Safety of Trametinib in Neurofibromatosis Type 1-Associated Plexiform Neurofibroma and Low-Grade Glioma: A Systematic Review and Meta-Analysis. *Pharmaceuticals* 2022, 15, 956. [CrossRef]
- 136. Farazdaghi, M.K.; Katowitz, W.R.; Avery, R.A. Current Treatment of Optic Nerve Gliomas. *Curr. Opin. Ophthalmol.* 2019, 30, 356–363. [CrossRef]
- 137. Bataini, J.P.; Delanian, S.; Ponvert, D. Chiasmal Gliomas: Results of Irradiation Management in 57 Patients and Review of Literature. *Int. J. Radiat. Oncol. Biol. Phys.* **1991**, *21*, 615–623. [CrossRef]
- Cappelli, C.; Grill, J.; Raquin, M.; Pierre-Kahn, A.; Lellouch-Tubiana, A.; Terrier-Lacombe, M.J.; Habrand, J.L.; Couanet, D.; Brauner, R.; Rodriguez, D.; et al. Long-Term Follow up of 69 Patients Treated for Optic Pathway Tumours before the Chemotherapy Era. Arch. Dis. Child 1998, 79, 334–338. [CrossRef]
- Lacaze, E.; Kieffer, V.; Streri, A.; Lorenzi, C.; Gentaz, E.; Habrand, J.-L.; Dellatolas, G.; Kalifa, C.; Grill, J. Neuropsychological Outcome in Children with Optic Pathway Tumours When First-Line Treatment Is Chemotherapy. *Br. J. Cancer* 2003, *89*, 2038–2044. [CrossRef]
- Sutton, L.N.; Molloy, P.T.; Sernyak, H.; Goldwein, J.; Phillips, P.L.; Rorke, L.B.; Moshang, T.; Lange, B.; Packer, R.J. Long-Term Outcome of Hypothalamic/Chiasmatic Astrocytomas in Children Treated with Conservative Surgery. *J. Neurosurg.* 1995, 83, 583–589. [CrossRef]
- 141. Rosser, T.L.; Vezina, G.; Packer, R.J. Cerebrovascular Abnormalities in a Population of Children with Neurofibromatosis Type 1. *Neurology* **2005**, *64*, 553–555. [CrossRef]
- 142. Ullrich, N.J.; Robertson, R.; Kinnamon, D.D.; Scott, R.M.; Kieran, M.W.; Turner, C.D.; Chi, S.N.; Goumnerova, L.; Proctor, M.; Tarbell, N.J.; et al. Moyamoya Following Cranial Irradiation for Primary Brain Tumors in Children. *Neurology* 2007, 68, 932–938. [CrossRef]
- 143. Grabenbauer, G.G.; Schuchardt, U.; Buchfelder, M.; Rödel, C.M.; Gusek, G.; Marx, M.; Doerr, H.G.; Fahlbusch, R.; Huk, W.J.; Wenzel, D.; et al. Radiation Therapy of Optico-Hypothalamic Gliomas (OHG)--Radiographic Response, Vision and Late Toxicity. *Radiother. Oncol.* 2000, 54, 239–245. [CrossRef]
- 144. Horwich, A.; Bloom, H.J. Optic Gliomas: Radiation Therapy and Prognosis. *Int. J. Radiat. Oncol. Biol. Phys.* **1985**, *11*, 1067–1079. [CrossRef]
- 145. Awdeh, R.M.; Kiehna, E.N.; Drewry, R.D.; Kerr, N.C.; Haik, B.G.; Wu, S.; Xiong, X.; Merchant, T.E. Visual Outcomes in Pediatric Optic Pathway Glioma after Conformal Radiation Therapy. *Int. J. Radiat. Oncol. Biol. Phys.* **2012**, *84*, 46–51. [CrossRef]
- El-Shehaby, A.M.N.; Reda, W.A.; Abdel Karim, K.M.; Emad Eldin, R.M.; Nabeel, A.M. Single-Session Gamma Knife Radiosurgery for Optic Pathway/Hypothalamic Gliomas. J. Neurosurg. 2016, 125, 50–57. [CrossRef]
- 147. Dong, M.-J.; Yang, Z.-K.; Yang, J.; Guo, R.-Q.; Xiao, Y.-Y.; Liu, H. Gamma Knife Radiotherapy in a Neurofibromatosis Type 1 Chinese Pedigrees with NF1 Gene Frameshift Mutation: A Case Report. *Medicine* 2022, 101, e29280. [CrossRef]
- 148. Saran, F.H.; Baumert, B.G.; Khoo, V.S.; Adams, E.J.; Garré, M.L.; Warrington, A.P.; Brada, M. Stereotactically Guided Conformal Radiotherapy for Progressive Low-Grade Gliomas of Childhood. *Int. J. Radiat. Oncol. Biol. Phys.* **2002**, *53*, 43–51. [CrossRef]
- 149. Combs, S.E.; Schulz-Ertner, D.; Moschos, D.; Thilmann, C.; Huber, P.E.; Debus, J. Fractionated Stereotactic Radiotherapy of Optic Pathway Gliomas: Tolerance and Long-Term Outcome. *Int. J. Radiat. Oncol. Biol. Phys.* **2005**, *62*, 814–819. [CrossRef]
- 150. Hug, E.B.; Muenter, M.W.; Archambeau, J.O.; DeVries, A.; Liwnicz, B.; Loredo, L.N.; Grove, R.I.; Slater, J.D. Conformal Proton Radiation Therapy for Pediatric Low-Grade Astrocytomas. *Strahlenther. Onkol.* **2002**, *178*, 10–17. [CrossRef]
- 151. Thirunavu, V.M.; Mohammad, L.M.; Kandula, V.; Beestrum, M.; Lam, S.K. Vision Outcomes for Pediatric Patients with Optic Pathway Gliomas Associated with Neurofibromatosis Type I: A Systematic Review of the Clinical Evidence. *J. Pediatr. Hematol. Oncol.* **2021**, *43*, 135–143. [CrossRef]
- Packer, R.J.; Ater, J.; Allen, J.; Phillips, P.; Geyer, R.; Nicholson, H.S.; Jakacki, R.; Kurczynski, E.; Needle, M.; Finlay, J.; et al. Carboplatin and Vincristine Chemotherapy for Children with Newly Diagnosed Progressive Low-Grade Gliomas. *J. Neurosurg.* 1997, *86*, 747–754. [CrossRef]
- 153. Lafay-Cousin, L.; Holm, S.; Qaddoumi, I.; Nicolin, G.; Bartels, U.; Tabori, U.; Huang, A.; Bouffet, E. Weekly Vinblastine in Pediatric Low-Grade Glioma Patients with Carboplatin Allergic Reaction. *Cancer* **2005**, *103*, 2636–2642. [CrossRef]
- 154. Massimino, M.; Spreafico, F.; Cefalo, G.; Riccardi, R.; Tesoro-Tess, J.D.; Gandola, L.; Riva, D.; Ruggiero, A.; Valentini, L.; Mazza, E.; et al. High Response Rate to Cisplatin/Etoposide Regimen in Childhood Low-Grade Glioma. *J. Clin. Oncol.* 2002, 20, 4209–4216. [CrossRef]
- 155. Ater, J.L.; Zhou, T.; Holmes, E.; Mazewski, C.M.; Booth, T.N.; Freyer, D.R.; Lazarus, K.H.; Packer, R.J.; Prados, M.; Sposto, R.; et al. Randomized Study of Two Chemotherapy Regimens for Treatment of Low-Grade Glioma in Young Children: A Report from the Children's Oncology Group. J. Clin. Oncol. 2012, 30, 2641–2647. [CrossRef]
- 156. Felix, C.A. Secondary Leukemias Induced by Topoisomerase-Targeted Drugs. *Biochim. Biophys. Acta* 1998, 1400, 233–255. [CrossRef]
- 157. Perry, J.R.; Brown, M.T.; Gockerman, J.P. Acute Leukemia Following Treatment of Malignant Glioma. *J. Neuro-Oncol.* **1998**, 40, 39–46. [CrossRef]

- 158. Bouffet, E.; Jakacki, R.; Goldman, S.; Hargrave, D.; Hawkins, C.; Shroff, M.; Hukin, J.; Bartels, U.; Foreman, N.; Kellie, S.; et al. Phase II Study of Weekly Vinblastine in Recurrent or Refractory Pediatric Low-Grade Glioma. J. Clin. Oncol. 2012, 30, 1358–1363. [CrossRef]
- 159. Lassaletta, A.; Scheinemann, K.; Zelcer, S.M.; Hukin, J.; Wilson, B.A.; Jabado, N.; Carret, A.S.; Lafay-Cousin, L.; Larouche, V.; Hawkins, C.E.; et al. Phase II Weekly Vinblastine for Chemotherapy-Naïve Children with Progressive Low-Grade Glioma: A Canadian Pediatric Brain Tumor Consortium Study. J. Clin. Oncol. 2016, 34, 3537–3543. [CrossRef]
- 160. Cappellano, A.M.; Petrilli, A.S.; da Silva, N.S.; Silva, F.A.; Paiva, P.M.; Cavalheiro, S.; Bouffet, E. Single Agent Vinorelbine in Pediatric Patients with Progressive Optic Pathway Glioma. *J. Neurooncol.* **2015**, *121*, 405–412. [CrossRef]
- 161. Wisinski, K.B.; Flamand, Y.; Wilson, M.A.; Luke, J.J.; Tawbi, H.A.; Hong, F.; Mitchell, E.P.; Zwiebel, J.A.; Chen, H.; Gray, R.J.; et al. Trametinib in Patients with NF1-, GNAQ-, or GNA11-Mutant Tumors: Results from the NCI-MATCH ECOG-ACRIN Trial (EAY131) Subprotocols S1 and S2. JCO Precis. Oncol. 2023, 7, e2200421. [CrossRef]
- 162. Gururangan, S.; Fisher, M.J.; Allen, J.C.; Herndon, J.E.; Quinn, J.A.; Reardon, D.A.; Vredenburgh, J.J.; Desjardins, A.; Phillips, P.C.; Watral, M.A.; et al. Temozolomide in Children with Progressive Low-Grade Glioma. *Neuro-Oncology* **2007**, *9*, 161–168. [CrossRef]
- 163. De Vita, S.; De Matteis, S.; Laurenti, L.; Chiusolo, P.; Reddiconto, G.; Fiorini, A.; Leone, G.; Sica, S. Secondary Ph+ Acute Lymphoblastic Leukemia after Temozolomide. *Ann. Hematol.* **2005**, *84*, 760–762. [CrossRef]
- 164. Shofty, B.; Ben-Sira, L.; Kesler, A.; Jallo, G.; Groves, M.L.; Iyer, R.R.; Lassaletta, A.; Tabori, U.; Bouffet, E.; Thomale, U.-W.; et al. Isolated Optic Nerve Gliomas: A Multicenter Historical Cohort Study. J. Neurosurg. Pediatr. 2017, 20, 549–555. [CrossRef]
- 165. Listernick, R.; Ferner, R.E.; Piersall, L.; Sharif, S.; Gutmann, D.H.; Charrow, J. Late-Onset Optic Pathway Tumors in Children with Neurofibromatosis 1. *Neurology* **2004**, *63*, 1944–1946. [CrossRef]
- Presta, L.G.; Chen, H.; O'Connor, S.J.; Chisholm, V.; Meng, Y.G.; Krummen, L.; Winkler, M.; Ferrara, N. Humanization of an Anti-Vascular Endothelial Growth Factor Monoclonal Antibody for the Therapy of Solid Tumors and Other Disorders. *Cancer Res.* 1997, 57, 4593–4599.
- 167. Hwang, E.I.; Jakacki, R.I.; Fisher, M.J.; Kilburn, L.B.; Horn, M.; Vezina, G.; Rood, B.R.; Packer, R.J. Long-Term Efficacy and Toxicity of Bevacizumab-Based Therapy in Children with Recurrent Low-Grade Gliomas. *Pediatr. Blood Cancer* 2013, 60, 776–782. [CrossRef]
- 168. Kalra, M.; Heath, J.A.; Kellie, S.J.; Dalla Pozza, L.; Stevens, M.M.; Swamy, S.; McCowage, G.B. Confirmation of Bevacizumab Activity, and Maintenance of Efficacy in Retreatment after Subsequent Relapse, in Pediatric Low-Grade Glioma. *J. Pediatr. Hematol. Oncol.* **2015**, *37*, e341–e346. [CrossRef]
- Packer, R.J.; Jakacki, R.; Horn, M.; Rood, B.; Vezina, G.; MacDonald, T.; Fisher, M.J.; Cohen, B. Objective Response of Multiply Recurrent Low-Grade Gliomas to Bevacizumab and Irinotecan. *Pediatr. Blood Cancer* 2009, 52, 791–795. [CrossRef]
- Couec, M.-L.; André, N.; Thebaud, E.; Minckes, O.; Rialland, X.; Corradini, N.; Aerts, I.; Marec Bérard, P.; Bourdeaut, F.; Leblond, P.; et al. Bevacizumab and Irinotecan in Children with Recurrent or Refractory Brain Tumors: Toxicity and Efficacy Trends. *Pediatr. Blood Cancer* 2012, 59, 34–38. [CrossRef]
- 171. Zhukova, N.; Rajagopal, R.; Lam, A.; Coleman, L.; Shipman, P.; Walwyn, T.; Williams, M.; Sullivan, M.; Campbell, M.; Bhatia, K.; et al. Use of Bevacizumab as a Single Agent or in Adjunct with Traditional Chemotherapy Regimens in Children with Unresectable or Progressive Low-Grade Glioma. *Cancer Med.* 2019, *8*, 40–50. [CrossRef]
- 172. Green, K.; Panagopoulou, P.; D'Arco, F.; O'Hare, P.; Bowman, R.; Walters, B.; Dahl, C.; Jorgensen, M.; Patel, P.; Slater, O.; et al. A Nationwide Evaluation of Bevacizumab-Based Treatments in Pediatric Low-Grade Glioma in the UK: Safety, Efficacy, Visual Morbidity, and Outcomes. *Neuro-Oncology* 2023, 25, 774–785. [CrossRef]
- 173. Yamasaki, F.; Takano, M.; Yonezawa, U.; Taguchi, A.; Kolakshyapati, M.; Okumichi, H.; Kiuchi, Y.; Kurisu, K. Bevacizumab for Optic Pathway Glioma with Worsening Visual Field in Absence of Imaging Progression: 2 Case Reports and Literature Review. *Childs Nerv. Syst.* **2020**, *36*, 635–639. [CrossRef]
- 174. Fangusaro, J.; Onar-Thomas, A.; Young Poussaint, T.; Wu, S.; Ligon, A.H.; Lindeman, N.; Banerjee, A.; Packer, R.J.; Kilburn, L.B.; Goldman, S.; et al. Selumetinib in Paediatric Patients with BRAF-Aberrant or Neurofibromatosis Type 1-Associated Recurrent, Refractory, or Progressive Low-Grade Glioma: A Multicentre, Phase 2 Trial. *Lancet Oncol.* **2019**, *20*, 1011–1022. [CrossRef]
- 175. Walsh, K.S.; Wolters, P.L.; Widemann, B.C.; Del Castillo, A.; Sady, M.D.; Inker, T.; Roderick, M.C.; Martin, S.; Toledo-Tamula, M.A.; Struemph, K.; et al. Impact of MEK Inhibitor Therapy on Neurocognitive Functioning in NF1. *Neurol. Genet.* 2021, 7, e616. [CrossRef]
- 176. Harder, A. MEK Inhibitors-Novel Targeted Therapies of Neurofibromatosis Associated Benign and Malignant Lesions. *Biomark. Res.* **2021**, *9*, 26. [CrossRef]
- 177. Pillay-Smiley, N.; Fletcher, J.S.; de Blank, P.; Ratner, N. Shedding New Light: Novel Therapies for Common Disorders in Children with Neurofibromatosis Type I. *Pediatr. Clin. N. Am.* 2023, 70, 937–950. [CrossRef]
- 178. Del Bufalo, F.; Ceglie, G.; Cacchione, A.; Alessi, I.; Colafati, G.S.; Carai, A.; Diomedi-Camassei, F.; De Billy, E.; Agolini, E.; Mastronuzzi, A.; et al. BRAF V600E Inhibitor (Vemurafenib) for BRAF V600E Mutated Low Grade Gliomas. *Front. Oncol.* 2018, 8, 526. [CrossRef]
- 179. Ullrich, N.J.; Prabhu, S.P.; Packer, R.J.; Goldman, S.; Robison, N.J.; Allen, J.C.; Viskochil, D.H.; Gutmann, D.H.; Perentesis, J.P.; Korf, B.R.; et al. Visual Outcomes Following Everolimus Targeted Therapy for Neurofibromatosis Type 1-Associated Optic Pathway Gliomas in Children. *Pediatr. Blood Cancer* **2021**, *68*, e28833. [CrossRef]

- Ullrich, N.J.; Prabhu, S.P.; Reddy, A.T.; Fisher, M.J.; Packer, R.; Goldman, S.; Robison, N.J.; Gutmann, D.H.; Viskochil, D.H.; Allen, J.C.; et al. A Phase II Study of Continuous Oral MTOR Inhibitor Everolimus for Recurrent, Radiographic-Progressive Neurofibromatosis Type 1-Associated Pediatric Low-Grade Glioma: A Neurofibromatosis Clinical Trials Consortium Study. *Neuro-Oncology* 2020, 22, 1527–1535. [CrossRef]
- 181. Falsini, B.; Chiaretti, A.; Rizzo, D.; Piccardi, M.; Ruggiero, A.; Manni, L.; Soligo, M.; Dickmann, A.; Federici, M.; Salerni, A.; et al. Nerve Growth Factor Improves Visual Loss in Childhood Optic Gliomas: A Randomized, Double-Blind, Phase II Clinical Trial. *Brain* 2016, 139, 404–414. [CrossRef]
- 182. Brannan, C.I.; Perkins, A.S.; Vogel, K.S.; Ratner, N.; Nordlund, M.L.; Reid, S.W.; Buchberg, A.M.; Jenkins, N.A.; Parada, L.F.; Copeland, N.G. Targeted Disruption of the Neurofibromatosis Type-1 Gene Leads to Developmental Abnormalities in Heart and Various Neural Crest-Derived Tissues. *Genes Dev.* 1994, *8*, 1019–1029. [CrossRef]
- Jacks, T.; Shih, T.S.; Schmitt, E.M.; Bronson, R.T.; Bernards, A.; Weinberg, R.A. Tumour Predisposition in Mice Heterozygous for a Targeted Mutation in Nf1. *Nat. Genet.* 1994, 7, 353–361. [CrossRef]
- Toonen, J.A.; Anastasaki, C.; Smithson, L.J.; Gianino, S.M.; Li, K.; Kesterson, R.A.; Gutmann, D.H. NF1 Germline Mutation Differentially Dictates Optic Glioma Formation and Growth in Neurofibromatosis-1. *Hum. Mol. Genet.* 2016, 25, 1703–1713. [CrossRef]
- 185. Zhu, Y.; Romero, M.I.; Ghosh, P.; Ye, Z.; Charnay, P.; Rushing, E.J.; Marth, J.D.; Parada, L.F. Ablation of NF1 Function in Neurons Induces Abnormal Development of Cerebral Cortex and Reactive Gliosis in the Brain. *Genes Dev.* **2001**, *15*, 859–876. [CrossRef]
- 186. Bajenaru, M.L.; Hernandez, M.R.; Perry, A.; Zhu, Y.; Parada, L.F.; Garbow, J.R.; Gutmann, D.H. Optic Nerve Glioma in Mice Requires Astrocyte Nf1 Gene Inactivation and Nf1 Brain Heterozygosity. *Cancer Res.* 2003, 63, 8573–8577.
- 187. Zhu, Y.; Harada, T.; Liu, L.; Lush, M.E.; Guignard, F.; Harada, C.; Burns, D.K.; Bajenaru, M.L.; Gutmann, D.H.; Parada, L.F. Inactivation of NF1 in CNS Causes Increased Glial Progenitor Proliferation and Optic Glioma Formation. *Development* 2005, 132, 5577–5588. [CrossRef]
- 188. Yvone, G.M.; Breunig, J.J. Pediatric Low-Grade Glioma Models: Advances and Ongoing Challenges. *Front. Oncol.* 2023, 13, 1346949. [CrossRef]
- Dasgupta, B.; Li, W.; Perry, A.; Gutmann, D.H. Glioma Formation in Neurofibromatosis 1 Reflects Preferential Activation of K-RAS in Astrocytes. *Cancer Res.* 2005, 65, 236–245. [CrossRef]
- 190. Hegedus, B.; Dasgupta, B.; Shin, J.E.; Emnett, R.J.; Hart-Mahon, E.K.; Elghazi, L.; Bernal-Mizrachi, E.; Gutmann, D.H. Neurofibromatosis-1 Regulates Neuronal and Glial Cell Differentiation from Neuroglial Progenitors in Vivo by Both CAMP- and Ras-Dependent Mechanisms. *Cell Stem Cell* 2007, 1, 443–457. [CrossRef]
- 191. Solga, A.C.; Toonen, J.A.; Pan, Y.; Cimino, P.J.; Ma, Y.; Castillon, G.A.; Gianino, S.M.; Ellisman, M.H.; Lee, D.Y.; Gutmann, D.H. The Cell of Origin Dictates the Temporal Course of Neurofibromatosis-1 (Nf1) Low-Grade Glioma Formation. *Oncotarget* 2017, *8*, 47206–47215. [CrossRef]
- 192. Chatterjee, J.; Koleske, J.P.; Chao, A.; Sauerbeck, A.D.; Chen, J.-K.; Qi, X.; Ouyang, M.; Boggs, L.G.; Idate, R.; Marco Y Marquez, L.I.; et al. Brain Injury Drives Optic Glioma Formation through Neuron-Glia Signaling. *Acta Neuropathol. Commun.* 2024, 12, 21. [CrossRef]
- 193. Banerjee, S.; Crouse, N.R.; Emnett, R.J.; Gianino, S.M.; Gutmann, D.H. Neurofibromatosis-1 Regulates MTOR-Mediated Astrocyte Growth and Glioma Formation in a TSC/Rheb-Independent Manner. *Proc. Natl. Acad. Sci. USA* 2011, 108, 15996–16001. [CrossRef]
- Daginakatte, G.C.; Gianino, S.M.; Zhao, N.W.; Parsadanian, A.S.; Gutmann, D.H. Increased C-Jun-NH2-Kinase Signaling in Neurofibromatosis-1 Heterozygous Microglia Drives Microglia Activation and Promotes Optic Glioma Proliferation. *Cancer Res.* 2008, 68, 10358–10366. [CrossRef]
- 195. Pong, W.W.; Higer, S.B.; Gianino, S.M.; Emnett, R.J.; Gutmann, D.H. Reduced Microglial CX3CR1 Expression Delays Neurofibromatosis-1 Glioma Formation. *Ann. Neurol.* 2013, 73, 303–308. [CrossRef]
- Logiacco, F.; Grzegorzek, L.C.; Cordell, E.C.; Popp, O.; Mertins, P.; Gutmann, D.H.; Kettenmann, H.; Semtner, M. Neurofibromatosis Type 1-Dependent Alterations in Mouse Microglia Function Are Not Cell-Intrinsic. *Acta Neuropathol. Commun.* 2023, 11, 36. [CrossRef]
- 197. Guo, X.; Pan, Y.; Xiong, M.; Sanapala, S.; Anastasaki, C.; Cobb, O.; Dahiya, S.; Gutmann, D.H. Midkine Activation of CD8+ T Cells Establishes a Neuron-Immune-Cancer Axis Responsible for Low-Grade Glioma Growth. *Nat. Commun.* 2020, *11*, 2177. [CrossRef]
- 198. Lee, D.Y.; Gianino, S.M.; Gutmann, D.H. Innate Neural Stem Cell Heterogeneity Determines the Patterning of Glioma Formation in Children. *Cancer Cell* **2012**, *22*, 131–138. [CrossRef]
- 199. Chen, Y.-H.; McGowan, L.D.; Cimino, P.J.; Dahiya, S.; Leonard, J.R.; Lee, D.Y.; Gutmann, D.H. Mouse Low-Grade Gliomas Contain Cancer Stem Cells with Unique Molecular and Functional Properties. *Cell Rep.* **2015**, *10*, 1899–1912. [CrossRef]
- Wang, Y.; Kim, E.; Wang, X.; Novitch, B.G.; Yoshikawa, K.; Chang, L.-S.; Zhu, Y. ERK Inhibition Rescues Defects in Fate Specification of Nf1-Deficient Neural Progenitors and Brain Abnormalities. *Cell* 2012, 150, 816–830. [CrossRef]
- 201. Jecrois, E.S.; Zheng, W.; Bornhorst, M.; Li, Y.; Treisman, D.M.; Muguyo, D.; Huynh, S.; Andrew, S.F.; Wang, Y.; Jiang, J.; et al. Treatment during a Developmental Window Prevents NF1-Associated Optic Pathway Gliomas by Targeting Erk-Dependent Migrating Glial Progenitors. Dev. Cell 2021, 56, 2871–2885.e6. [CrossRef]
- 202. Tang, Y.; Chatterjee, J.; Wagoner, N.; Bozeman, S.; Gutmann, D.H. Estrogen-Induced Glial IL-1β Mediates Extrinsic Retinal Ganglion Cell Vulnerability in Murine Nf1 Optic Glioma. *Ann. Clin. Transl. Neurol.* 2024, 11, 812–818. [CrossRef]

- 203. Osum, S.H.; Watson, A.L.; Largaespada, D.A. Spontaneous and Engineered Large Animal Models of Neurofibromatosis Type 1. *Int. J. Mol. Sci.* 2021, 22, 1954. [CrossRef]
- Métin, C.; Irons, W.A.; Frost, D.O. Retinal Ganglion Cells in Normal Hamsters and Hamsters with Novel Retinal Projections. I. Number, Distribution, and Size. J. Comp. Neurol. 1995, 353, 179–199. [CrossRef] [PubMed]
- 205. Nadal-Nicolás, F.M.; Jiménez-López, M.; Salinas-Navarro, M.; Sobrado-Calvo, P.; Alburquerque-Béjar, J.J.; Vidal-Sanz, M.; Agudo-Barriuso, M. Whole Number, Distribution and Co-Expression of Brn3 Transcription Factors in Retinal Ganglion Cells of Adult Albino and Pigmented Rats. PLoS ONE 2012, 7, e49830. [CrossRef] [PubMed]
- Nadal-Nicolás, F.M.; Salinas-Navarro, M.; Jiménez-López, M.; Sobrado-Calvo, P.; Villegas-Pérez, M.P.; Vidal-Sanz, M.; Agudo-Barriuso, M. Displaced Retinal Ganglion Cells in Albino and Pigmented Rats. Front. Neuroanat. 2014, 8, 99. [CrossRef]
- 207. Isakson, S.H.; Rizzardi, A.E.; Coutts, A.W.; Carlson, D.F.; Kirstein, M.N.; Fisher, J.; Vitte, J.; Williams, K.B.; Pluhar, G.E.; Dahiya, S.; et al. Genetically Engineered Minipigs Model the Major Clinical Features of Human Neurofibromatosis Type 1. *Commun. Biol.* 2018, 1, 158. [CrossRef]
- 208. White, K.A.; Swier, V.J.; Cain, J.T.; Kohlmeyer, J.L.; Meyerholz, D.K.; Tanas, M.R.; Uthoff, J.; Hammond, E.; Li, H.; Rohret, F.A.; et al. A Porcine Model of Neurofibromatosis Type 1 That Mimics the Human Disease. JCI Insight 2018, 3, e120402. [CrossRef]
- Monlux, A.W.; Davis, C.L. Multiple Schwannomas of Cattle (Nerve Sheath Tumors; Multiple Neurilemmomas; Neurofibromatosis). Am. J. Vet. Res. 1953, 14, 499–509.
- 210. Schöniger, S.; Summers, B.A. Localized, Plexiform, Diffuse, and Other Variants of Neurofibroma in 12 Dogs, 2 Horses, and a Chicken. *Vet. Pathol.* 2009, 46, 904–915. [CrossRef]
- Miettinen, M.M.; Antonescu, C.R.; Fletcher, C.D.M.; Kim, A.; Lazar, A.J.; Quezado, M.M.; Reilly, K.M.; Stemmer-Rachamimov, A.; Stewart, D.R.; Viskochil, D.; et al. Histopathologic Evaluation of Atypical Neurofibromatous Tumors and Their Transformation into Malignant Peripheral Nerve Sheath Tumor in Patients with Neurofibromatosis 1-a Consensus Overview. *Hum. Pathol.* 2017, 67, 1–10. [CrossRef]
- 212. Shin, J.; Padmanabhan, A.; de Groh, E.D.; Lee, J.-S.; Haidar, S.; Dahlberg, S.; Guo, F.; He, S.; Wolman, M.A.; Granato, M.; et al. Zebrafish Neurofibromatosis Type 1 Genes Have Redundant Functions in Tumorigenesis and Embryonic Development. *Dis. Model Mech.* 2012, *5*, 881–894. [CrossRef]
- 213. He, S.; Mansour, M.R.; Zimmerman, M.W.; Ki, D.H.; Layden, H.M.; Akahane, K.; Gjini, E.; de Groh, E.D.; Perez-Atayde, A.R.; Zhu, S.; et al. Synergy between Loss of NF1 and Overexpression of MYCN in Neuroblastoma Is Mediated by the GAP-Related Domain. *eLife* 2016, *5*, e14713. [CrossRef]
- 214. King, L.B.; Boto, T.; Botero, V.; Aviles, A.M.; Jomsky, B.M.; Joseph, C.; Walker, J.A.; Tomchik, S.M. Developmental Loss of Neurofibromin across Distributed Neuronal Circuits Drives Excessive Grooming in Drosophila. *PLoS Genet.* 2020, 16, e1008920. [CrossRef] [PubMed]
- 215. The, I.; Hannigan, G.E.; Cowley, G.S.; Reginald, S.; Zhong, Y.; Gusella, J.F.; Hariharan, I.K.; Bernards, A. Rescue of a Drosophila NF1 Mutant Phenotype by Protein Kinase A. *Science* 1997, 276, 791–794. [CrossRef] [PubMed]
- 216. Yuan, M.; White, D.; Resar, L.; Bar, E.; Groves, M.; Cohen, A.; Jackson, E.; Bynum, J.; Rubens, J.; Mumm, J.; et al. Conditional Reprogramming Culture Conditions Facilitate Growth of Lower-Grade Glioma Models. *Neuro-Oncology* 2021, 23, 770–782. [CrossRef] [PubMed]
- 217. Boia, R.; Ruzafa, N.; Aires, I.D.; Pereiro, X.; Ambrósio, A.F.; Vecino, E.; Santiago, A.R. Neuroprotective Strategies for Retinal Ganglion Cell Degeneration: Current Status and Challenges Ahead. *Int. J. Mol. Sci.* **2020**, *21*, 2262. [CrossRef] [PubMed]
- Auricchio, A.; Smith, A.J.; Ali, R.R. The Future Looks Brighter After 25 Years of Retinal Gene Therapy. *Hum. Gene Ther.* 2017, 28, 982–987. [CrossRef] [PubMed]
- Luo, Z.; Chang, K.-C.; Wu, S.; Sun, C.; Xia, X.; Nahmou, M.; Bian, M.; Wen, R.R.; Zhu, Y.; Shah, S.; et al. Directly Induced Human Retinal Ganglion Cells Mimic Fetal RGCs and Are Neuroprotective after Transplantation in Vivo. *Stem Cell Rep.* 2022, 17, 2690–2703. [CrossRef]
- 220. Millán-Rivero, J.E.; Nadal-Nicolás, F.M.; García-Bernal, D.; Sobrado-Calvo, P.; Blanquer, M.; Moraleda, J.M.; Vidal-Sanz, M.; Agudo-Barriuso, M. Human Wharton's Jelly Mesenchymal Stem Cells Protect Axotomized Rat Retinal Ganglion Cells via Secretion of Anti-Inflammatory and Neurotrophic Factors. *Sci. Rep.* 2018, *8*, 16299. [CrossRef]

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.