

Review

A Critical Review on In Vitro and Ex Vivo Models of the Intestinal Epithelium of Humans and Monogastric Animals

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Abstract: Recently, the bioactive potential of several functional ingredients and biomolecules has been evaluated regarding human and animal nutrition. The digestive process from food intake to absorption and metabolism are important events that induce changes in ingredients, which affect their bioactivity. Consequently, there is a need to assess the bioavailability and bioaccessibility of these compounds. The methodology for the simulation of the human gastrointestinal tract has been standardized (INFOGEST protocol), while a gastrointestinal protocol for other animals (e.g., ruminants or broilers) has yet to be established. However, INFOGEST allows us only to predict bioaccessibility, leaving a gap regarding a methodology able to assess bioavailability by mimicking intestinal permeability and absorption. Several approaches—including in vitro, ex vivo, in situ and in vivo methods—can be found in the literature, aiming to tackle transepithelial routes, but leading to different results concerning the bioefficiency of the compounds studied. Therefore, this review aims to assess the current state-of-the-art regarding monogastric intestinal dynamics, absorption, and permeability events. Moreover, it compiled methodologies for simulating intestinal absorption in several biological systems, while reasoning their advantages, disadvantages, applications in ingredient development and the existing gaps.

Keywords: ex vivo; in vitro; intestinal absorption; intestinal epithelium; monogastric animals



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1. Introduction

In recent decades, the role of nutrition has been widely recognized as a way to improve health and prevent a vast array of diseases in both humans and animals [1,2]. Therefore, in recent years, the bioactive potential of ingredients along with the development of so-called functional foods have been extensively explored by the scientific community. Bioefficiency is broadly defined as the potential of foods to fulfil the nutritional and metabolic needs of the consumer, so that the organism may perform its basic functions, but also contribute to health maintenance, through its bioactive potential [3]. However, the bioefficiency of many compounds with promising bioactivity may be affected by digestion, food intake, enzymatic digestion and absorption to metabolization. Hence, their bioefficiency should be clarified through the assessment of bioaccessibility (BC) and bioavailability (BA). Bioaccessibility usually refers to the fraction of compounds that are released as a result of gastrointestinal digestion and are therefore available to be absorbed [3,4]. As for BA, it stands for the fraction of compounds or metabolites that are absorbed unchanged, reach systemic circulation and then exert a bioactive effect on a target site, triggering a metabolic and/or physiologic response [4,5]. Thus, a compound, to fulfil its bioefficiency purpose, must first become bioaccessible, through digestion, so it can be absorbed into the systemic circulation and become bioavailable and finally exert its bioactive potential (Figure 1). In another way, the BC concept is focused on the impact of the digestive processes, including the stability

amidst acidic pH values or upon enzymatic action, while BA comprises the process of the intestinal absorption of the bioaccessible fraction, at the prompting of a physiological effect.

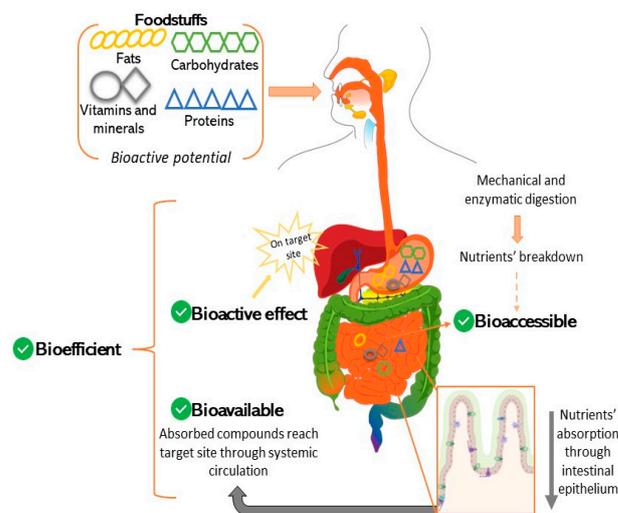


Figure 1. From food intake to bioefficiency—relationship between bioaccessibility, intestinal absorption, bioavailability and bioactivity.

Recently, several models of human and animal digestion simulation have been developed to predict the BC of different compounds. For example, INFOGEST is a widely acknowledged standardized static *in vitro* model of the human upper gastrointestinal tract (GIT), representing a valuable tool used to predict the BC of several ingredients [6,7]. Digestion simulation protocols have also been developed for chickens, since they have a distinct GIT among monogastric animals [8,9], despite not reuniting the same broad consensus as INFOGEST. From a different standpoint, BA represents a challenging concept to be accurately evaluated, since *in vivo* methodologies are the ‘gold standard’ and rely on measuring a compound of interest in blood plasma levels. However, there are currently several ethical constraints related, for example, with protecting animal’s welfare and human volunteers’ safety, in addition to being time and resource consuming [10–12]. As such, several models have been developed as attempts at predicting BA through the determination of the bioaccessible fraction absorbed. These systems are essentially tissue based, namely *ex vivo* (e.g., InTESTine™ Ussing chambers, Franz diffusion cells, everted-sac technique) or cell-culture based (e.g., Transwell® inserts, 3D organoids) [4,12,13]. Additionally, other alternatives are still being developed, such as continuous flow dialysis, organ chip technologies [14] and a medium throughput micro-physiological system, the intestinal Explant Barrier Chip (IEBC) set-up [15], among others.

From a different perspective, studies regarding the human GIT dynamics in health and disease demand more appropriate models, namely non-rodent ones, due to the intrinsic differences between the early stage GIT development in this species versus that of a human. As such, pig-based models have established themselves as the most reliable alternative supported by the extensive research on this animal and given the anatomical and physiological similarities, nutritional requirements and microbiota diversity [16,17]. Therefore, many studies on human health use pig tissues to assess the BA of compounds and drugs [18,19], surpassing the constraints of obtaining human intestinal segments. From a different perspective, models with other animal tissues can also be found in the literature, within the scope of animal science and nutrition. For instance, Ringø et al., 2014 [20] used salmon intestines in a Ussing chamber assay to assess the damaging effect of a pathogen, and Kent-Dennis et al., 2021 [21] used cow’s ruminal tissue for *ex vivo* studies regarding the effects of lipopolysaccharide (LPS), among many other studies. Additionally, the establishment of nutritional claims regarding ingredients’ bioactive properties through studies involving animal tissues are also being banished by several players that commercialize

functional ingredients, which opens the need for developing reliable permeation models based on artificial membranes.

The main purpose of this review was to address the lack of consensual and standardized methodologies and to assess the intestinal permeation of digested compounds by revising the existent models, as well as their adequacy, advantages and disadvantages. Moreover, it also intended to provide a comprehensive state-of-the-art review on the intestinal absorptive epithelium and its permeability. In addition to human and porcine absorption, given the acknowledged similarities, this review also compiled the most recent insights on chicken's intestinal absorption, in line with the body of work developed by our research group in recent years concerning the development of GIT in vitro models for humans and animals [22].

2. Materials and Methods

The research was carried out to assess the extent of the application of distinct techniques in simulating intestinal absorption. Google Scholar and PubMed were used with a time interval between 2015 and 2022. The search strings used combinations between animal model (pig, human, chicken), techniques (Ussing chamber, Franz diffusion, etc.) and other keywords regarding food matrices and intestinal permeability. For example: "pig" AND "Ussing chamber" AND "ingredient" AND "absorption".

Exclusion and Inclusion Criteria

The research papers were screened and selected based on whether their goal was to assess the effects of a nutrient/ingredient/supplement or molecule on health. Therefore, studies using different molecules (e.g., caffeine, among others) to compare distinct techniques and studies focused on the development/validation of new permeation models were excluded. In opposition, research papers not comprising GIT digestion before intestinal absorption simulation were included.

3. Overview of the Absorptive Epithelium—Structure and Physiology

The intestinal epithelium is a structure conserved among all vertebrates, serving the same purpose in distinct organism's physiology [23]. It consists of a sheet of tightly linked cells that separate the lumen from the internal environments. Moreover, it represents a primary site of absorption and establishes a constant interplay between the metabolism, immune system and microbiota [24]. Some key elements are common to several species, namely the presence of similar absorptive enterocytes, goblet cells, enteroendocrine cells or even the process of cellular turnover from the crypt to the top of the villus [25]. Additionally, these common threads and the availability of animal tissues turn animal models into valuable tools for exploring intestinal dynamics [25]. Therefore, this section intends to provide a basic understanding of the intestinal epitheliums of the models considered in this review.

3.1. Human Intestinal Epithelium

The human intestinal epithelium has been thoroughly reviewed and studied, and its involvement in health and disease has become clear. The reader can refer to complete reviews focused on this topic, such as [24,26,27]. While the complex mechanisms of cell structure and function and cell-to-cell communication are beyond the scope of this review, it is still relevant to keep in mind the basic notions of normal intestinal and epithelial organization (Figure 2A). The concepts of intestinal permeability and intestinal barrier have been explored as two distinct features of the same structure—the intestinal wall. On one hand, the term intestinal barrier refers to the protective aspect of the gut, acting as a physical barrier against microorganisms' invasion and preventing the passage of toxins. On the other hand, intestinal permeability relates to the transepithelial passage of fluids and nutrients [28]. When considering these two opposing characteristics, namely the provided protection, while allowing the absorption of fluids, it is important to understand how key structural

and physiological elements of the epithelium contribute to these characteristics (Figure 2B). From a different perspective, at a cellular level, the epithelial monolayer is sealed through three types of intercellular junctions: tight junctions (TJs), adherens junctions (AJs) and desmosomes. Each one of these junctions is in a specific membrane region contributing to, at a cellular level, the barrier function by safeguarding the paracellular pathway, but also contributing to the overall structural integrity and cell-to-cell communication. TJs represent the main controlling factor in the paracellular pathway, and their number increases from crypt to villus, along with the size selectivity. From another perspective, changes in the TJs' expression, often induced by proinflammatory cytokines, lipopolysaccharides (LPS) or pathogenic bacteria, have been linked to excessive intestinal permeability, which has been frequently associated with intestinal inflammation and diseases [29,30].

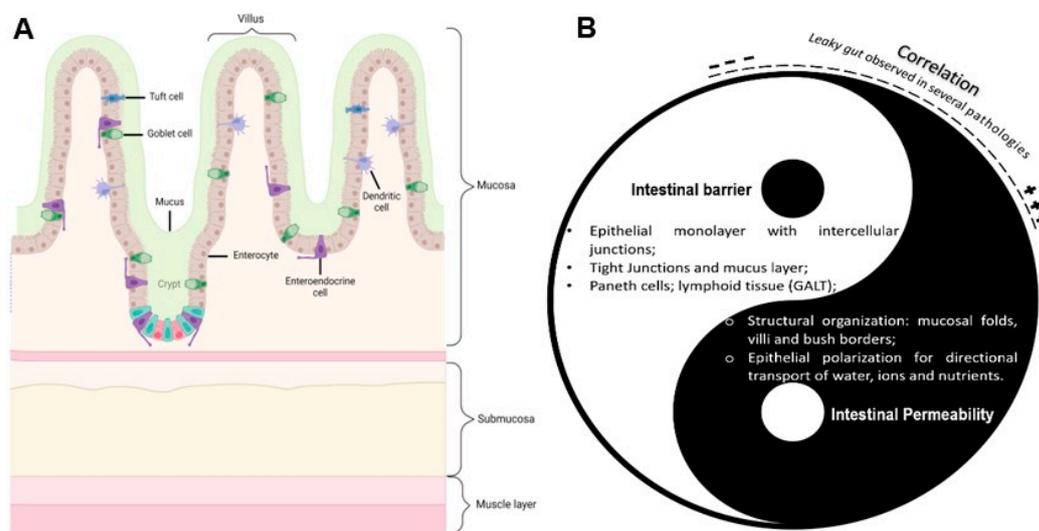


Figure 2. Intestinal epithelium structure and homeostasis. (A) Schematic representation of the key elements found in the small intestine epithelium and its main layers. Image created with Biorender.com. (B) The intestinal epithelium Yin–Yang: Intestinal permeability and intestinal barrier as two fundamental but opposite “forces” of the same structure. The symbols “+” and “–” express increases or decreases in gut permeability.

3.2. Pig and Chicken Intestinal Epitheliums

As mentioned above, a pig’s intestinal epithelium is very similar to that of a human, and therefore, it has also been extensively studied. Detailed reviews on the structural features of the intestinal epithelium can be found in Modina et al., 2019 and Modina et al., 2021 [31,32]. The epithelium is organized in four essential layers, as in most vertebrates. One macroscopic difference between human and pig intestines is regarding the intestinal length. The small intestine of a pig is ca. 20 m, that of a human averages around 6 m, and the pig’s large intestine has an average length of 5 m, while the human averages 1.5 m [33].

The chicken’s GIT has been described in the literature, in complete reviews such as those of Denbow, 2015 or Alshamy et al., 2018 [34,35]. Interestingly, the literature specifically targeting the intestinal epithelium and the absorption of nutrients is lacking. Briefly, the avian GIT is composed of an esophagus, crop, proventriculus, ventriculus/gizzard, small intestine, and large intestine. The crop is an out-pocket in the esophagus, where swallowed feed is stored until it passes on through the GIT. The proventriculus is the glandular stomach responsible for the secretion of acid and enzymes, as in chemical digestion. The gizzard or ventriculus, an organ unique in birds, functions as a mechanical stomach since it grinds, mixes and mashes the digesta and small stones. The small intestine is made up of the duodenum and the lower intestine, which is composed of the jejunum and ileum. The duodenum receives enzymes and bicarbonate from the pancreas and liver, which are particularly important for the digestion of proteins and lipids. Regarding nutrient

absorption, it occurs in the small intestine, through a specialized intestinal mucosa, with similar features to the ones described previously: villi and cellular polarization, with brush borders in the apical region to increase the absorption area [35]. The intestinal epithelium has a dynamic nature, in part influenced by nutrition since the villus height varies both with age and with nutritional intake [36,37].

3.3. Nutrients' Transportation Routes across Epithelium

Among monogastric organisms, the small intestine is usually the main site of nutrient absorption. The absorptive epithelial cells constitute ca. 90% of the epithelium, and their main function is to selectively uptake nutrients from the luminal environment, across the brush border. This is achieved through transepithelial transport, which is a dynamic and complex process, involving the modulation of the expressions of transmembrane transporting proteins, related to distinct transport mechanisms, but also varying according to the physicochemical attributes of the substances to be absorbed. Transport strategies can be essentially divided into paracellular transport—between two neighboring cells—and transcellular routes—across cells. The latter can be further subdivided into passive diffusion, carrier-mediated transport and endocytosis. The paracellular route is linked to the passive transport route, and it is mostly determined by the presence of TJs. The TJs permeability are mostly determined by the particle size. Therefore, the passage of substances through this route is minimal and essentially limited to solutes, water and ions, which cross the epithelium unaltered as they are not exposed to the intracellular environment [38]. Yu et al., 2016 [39] stated that the paracellular space, under normal physiological conditions, ranges from 0.3 to 1.0 nm. From a different perspective, the deregulation of the TJ complex has been linked to the serious impairment of the intestinal barrier (leaky gut) acting as a trigger for an inflammatory status and even systemic pathologies [40,41]. Transcellular transport is usually linked to the passage of proteins, macromolecules and larger particles.

As Kiela et al., 2016 [42] described, dietary sugars and amino acids are absorbed via facilitated diffusion, by transporters located in the brush border of the enterocytes. Glucose and galactose are absorbed via SGLT1 transporters, in a Na^+ -dependent manner (each sugar molecule is accompanied by two Na^+ ions), driven by an electrochemical gradient. Fructose is transported via GLUT5 and GLUT2, in the brush border and the basolateral membrane, respectively [43]. Di and tripeptides are absorbed through the H^+ /peptide transporter, but longer peptides (>tetrapeptides) are poorly absorbed. Some peptides are degraded into amino acids, through proteases. The amino acid transport is complex, given that countless systems exist that are responsible for amino acid uptake, which is generally ion dependent. Peptide transport is not dependent on the hydrophobicity of peptides, but essentially on peptide length, and their sequences seem to be a critical factor [44]. Lipids are essentially digested in the duodenum, by pancreatic enzymes and bile acids. Then, they are absorbed by the enterocytes via simple diffusion and move to the intracellular environment through the actions of CD36 and FABP (Fatty Acid-Binding Proteins) transporters [45]. Generally, only medium-length fatty acids can be absorbed. As for vitamins, for some of them, the transport mechanisms have not been fully disclosed (for example, in vitamins A and K) [46], and while some are absorbed through a transporter, others require more complex transport pathways since they often reach the intestine and require metabolic conversions.

Regarding swine's absorption mechanisms, they are similar to those of humans. The mechanisms for sugars, peptides and amino acid and lipid transport are the same [47,48]. For instance, fructose transporters are the same as in humans, belonging to the GLUT family. The only difference relates to the amino acid absorption in the large intestine, as previously observed in pigs, but not yet demonstrated in humans since it is generally accepted that amino acids that reach the large intestine are usually metabolized by the gut microbiota instead [49].

In chickens, the same basic mechanisms remain valid: sugars cross the epithelium through glucose transporters such as SGLT-1; the similar transporter applies to small peptide absorption (PepT-1), while the amino acid mechanism remains specific and com-

plex beyond the scope of the present review. Lipids are hydrolyzed into free fatty acids or monoglycerides and absorbed through simple diffusion, although other chaperones (FABP) are responsible for their intracellular movements [50]. Interestingly, Karasov, 2017 pointed out that birds have a higher paracellular absorption among flying and non-flying vertebrates [51].

Aside from the transport mechanisms described, it is also worth mentioning the close relationship between intestinal transport and the physicochemical properties of the compounds, namely their solubility (lipophilicity and/or hydrophilicity). Transport through passive diffusion is usually related to lipophilic compounds, meaning that compounds with a high affinity to the enterocyte's membrane, while hydrophilic compounds (water-soluble) cross the epithelium in a paracellular manner [42,52]. However, as Zhu et al., 2017 [53] stated, the paracellular route represents a minor pathway, as the intestinal surface area available for this route is reduced.

In summary, epithelial structure and physiology have been thoroughly described and studied in humans. However, despite humans' transepithelial nutrient transport mechanisms being well studied in terms of membrane transporters and drug absorption mechanisms, from a nutritional standpoint there are some knowledge gaps that remain to be addressed. For instance, as previously noted, the transport mechanisms of some vitamins are not completely disclosed. The void is even more evident in monogastric animals. The transepithelial absorptive routes in chickens are still poorly explored as information is lacking on this topic. The acknowledged link between animal health, its performance and its impact on human consumption accentuates the relevance of further investigating this topic [22].

4. In Vitro Models for Predicting Bioaccessibility

Although the focus of the present review is on models that simulate intestinal absorption, this process is included in the broader concept of digestion, and therefore, it is important to understand what models exist upstream of absorption. During digestion, nutrients present in the food/feed matrices are broken down through enzymatic and mechanical action and made bioaccessible for absorption. This process is essential for well-being as digestion can be considered an interface between foods and their impact on health [54]. In vivo methods consist of feeding animals or humans under controlled conditions. However, these models are expensive, complex and time consuming and increasingly raise ethical concerns. As such, developing better in vitro digestion models remains highly relevant when studying the impacts of bioactive compounds on health. The current models can be essentially divided into static models, meaning that only replicate biochemical processes, such as enzymatic action and pH value oscillations, and the products are immobilized, not simulating mechanical actions like mixing, shearing and others. Dynamic models contemplate pH changes, mechanical action, enzymatic secretions and the release of fluids through time, among others. Finally, semi-dynamic models combine features from both dynamic and static models. The complex features of the dynamic models, in opposition to the static ones, might translate into different bioaccessibilities, affecting the release of the food matrix, its solubility and stability. Dynamic models are more suited for kinetic studies, such as for determining nutrients' digestibility and for assessing nutrients' bioaccessibility, while static models are more appropriate for mechanist studies, since there are fewer variables to control and are more reproducible. Overall, in comparison to in vivo options, in vitro models are considered cheap and versatile, and their simplicity (especially of the static models) allows for standardization and comparisons with the literature. Additionally, they are particularly useful for screening the bioactive potential of new formulations/ingredients. However, gastrointestinal in vitro models are unable to mimic hormonal features, their interactions with the immune system or with the gut microbiota, for instance.

Currently, several models mimic human digestion, or parts of it. Overall, the INFOGEST models are widely used, and this consortium was able to significantly contribute

to the standardization of digestion protocols [6,55]. The INFOGEST protocol simulates almost the complete span of digestion, from the mouth to the small intestine. In each stage, conditions are standardized, concerning temperature, incubation time, pH value and fluid composition, and the enzymatic solutions are well defined. This protocol is very useful for either the kinetics of nutrient release or for monitoring the concentration or stability of a particular substance or nutrient. However, this model does not ensure complete fatty acid ionization in the small intestine step after lipid digestion and lacks dynamic feedback mechanisms. Nevertheless, studies have shown that despite these limitations, INFOGEST displays a good correlation with *in vivo* animal feeding studies. Other frequently cited *in vitro* models are the TNO Gastro-Intestinal Model (TIM), considering both the static and dynamic versions (TIM1 and TIM2), which simulate both stomach and intestinal digestion [56]; the Dynamic Gastrointestinal Simulator (GI Simulator—SIGMI) which is an automated *in vitro* gastrointestinal model specifically engineered to dynamically mimic the physiological events occurring during digestion, both in the stomach and small intestine. Additionally, it replicates the colonic microbiota, simulating its action within the large intestine [12]. SHIME is another unavoidable model since it mimics not only the stomach and the small intestine, but also the distinct regions of the colon [57].

As for animal *in vitro* models, there is a lack of standardization compared to that attained by INFOGEST. Regarding pig digestion, pigs have also been used as an animal model to perform *in vivo*–*in vitro* correlation studies for the INFOGEST protocol [58]. Additionally, INFOGEST has also been used in studies to predict amino acid digestion, given the similarities with the human gastrointestinal system [59]. The three-step method developed by Boisen et al., 1997 [60] represents a conventional approach to *in vitro* digestion simulation, with a general validity for the most used feedstuffs. This protocol contemplates proper particle size through grinding, incubation times, temperature and agitation, as well as pH variations, proper buffers and enzymatic solutions. These parameters are optimized for achieving the potential maximal digestibility. Most models mentioned in the literature aim to assess the digestibility of a particular nutrient, to better assess the energetic value of pig feeds. A more recent review on *in vitro* techniques for digestibility evaluation can be found in Świąch et al. (2017) [61], although the author also acknowledges the model proposed by Boisen as the most generally applicable one. Concerning chicken *in vitro* gastrointestinal models, the resources in the literature are even more scarce. Nash et al., 2022 [62] pointed out the over-reliance on *in vivo* studies in the face of the absence of proper *in vitro* models. The authors essentially reviewed cell culture approaches to drug development and toxicity studies, although acknowledging that the right models depend on the scientific question behind them. When considering the development of new functional feed formulations, there is a lack of complete and standardized models comprising the oral phase to the cecum simulation. Carvalho et al., (2023) proposed a model as a combination of several other models [63–65]. It includes mechanical action, incubation times/temperatures and enzymatic solutions for each digestive step and includes cecal fermentations, using inocula previously collected, covering to some extent the potential interactions of the ingredients in the study with the microbiota. This approach has the potential to be easily reproducible across different labs.

Overall, in what concerns human health, there are plenty of models, adequate for answering distinct questions. In what concerns the study of the bioactive potential of new ingredients, INFOGEST represents a crucial landmark in this field, reproducible and attainable to less resourceful research centers. In what concerns animal studies, particularly in the development of functional feedstuffs, there are models focused on assessing the digestibility of particular nutrients of energetic value, but there are no general or uniformized models for evaluating the potential impact of functional ingredients or additives on feeds. Additionally, in both humans and animals, there is a lack of studies that investigate how these upstream models of digestion could be combined with the downstream models focused in this review.

5. Intestinal Permeability Models and Applications

As previously stated, the BA depends on BC and absorption. While in vivo studies are the “gold standard” for assessing BA, they are laborious, bear high costs and pose ethical constraints that would be inadequate for the early screenings of ingredients or foodstuff development studies. The methodologies described are often grouped according to the underlying principle. Tissue-based methods usually refer to ex vivo procedures that rely on using an intestinal segment, for example, Franz diffusion cells, a Ussing chamber, or an everted gut sac. Other authors categorize according to biological or non-biological models, grouping cell-culture methods and ex vivo approaches, while differentiating them from nonbiological methods, such as artificial membranes. In the present review, the techniques described are classified into tissue-based methods, cell-culture-based methods, or other nonbiological methodologies that do not fit into those two major categories. Overall, ex vivo models can be considered a middle term between cell-based methods and non-biological models, concerning their physiological relevance. Since they maintain the tissue’s structure, including membrane transporters, nutrient absorption might take place through the active transport of passive diffusion mechanisms. Additionally, the integrity and structure of the epithelium are preserved. However, ex vivo models encompass a high tissue variability and lack a blood supply. Cell-based models display a wide range of complexity, from monolayer models to gut-on-chip models, which represent a promising technology. These models allow nutrient absorption through active transport or facilitated diffusion mechanisms. Furthermore, they do not have any ethical constraints and have a higher reproducibility. Non-biological methods usually have a higher throughput and are more cost-effective than ex vivo or cell-based methods, therefore being more suitable for initial screenings. Yet, they cannot mimic the complexity of the intestine. For instance, only the passive transport of nutrients has been considered [12,66]. The advantages and disadvantages of each model are also considered, as first systematized in Table 1, and further described in this section. Additionally, examples of their use in the ingredients of food formulation testing are also reviewed (Table 2).

Table 1. Main techniques and models for the assessment of intestinal absorption.

Technique	Description	Advantages	Disadvantages	
Tissue-based methods	Ussing chamber	Two-chamber system split up by a tissue segment in a horizontal orientation; two electrodes, one that provides current and another that measures TEER; continuous temperature and gas supply to media.	<ul style="list-style-type: none"> - Technique is well established and validated; - Allows for permeation evaluation across distinct intestinal regions; - Continuous system upgrade; - Allows the assessment of carrier-mediated transport. 	<ul style="list-style-type: none"> - Not adequate for all animal tissues; - Mandatory muscle layer removal; - High costs of system implementation and requires technical skills; - Low to medium throughput.
	Franz diffusion cells	System with two chambers (donor and receptor) displayed in a vertical orientation, divided by a tissue segment; controlled agitation and temperature.	<ul style="list-style-type: none"> - Less technically demanding than Ussing chamber; - Low-cost implementation system; - Versatile by allowing the use of distinct intestinal regions; - Medium throughput, as it depends on the number of Franz cells acquired. 	<ul style="list-style-type: none"> - Not robustly validated for intestinal permeation studies, as it is for skin permeation assays.
	Intestinal rings and segments	Isolated intestinal segment or ring submerged in a buffer solution with the compound of interest dissolved to assess its uptake through enterocytes.	<ul style="list-style-type: none"> - Practical and simple; - Agitation available (optional); - High throughput of samples; - Versatility that allows the evaluation of distinct intestinal sections; - More valuable for radiolabeled compounds. 	<ul style="list-style-type: none"> - Overly simplified system; - Does not allow for the determination of the direction of the transepithelial transport; - Decreasing relevance of this technique since less publications report it.
	Everted gut sac	Intestinal section reverted and filled with a buffer; both ends are tied, before incubation, with the compound in study; controlled temperature, aeration and stirring.	<ul style="list-style-type: none"> - Simple and inexpensive implementation; - High throughput; - Useful for determining drug absorption mechanisms and their metabolization and the roles of enzymes and transporters; - More useful if radiolabeled compounds are used. 	<ul style="list-style-type: none"> - Tissue reversion process may induce structural damages; - Leads to unrealistic absorption times.

Table 1. Cont.

Technique	Description	Advantages	Disadvantages
Cell-based methods	2D (monocultures/co-cultures of Caco-2 and/or HT29-MTX)	<ul style="list-style-type: none"> - Most traditional approach with several implemented protocols, with widely accepted results among regulatory institutions; - Highly proliferative cells that are relatively easy and cheap to culture; - Co-cultures of Caco-2 and HT29-MTX represent a more realistic approach than monocultures. 	<ul style="list-style-type: none"> - Reduced complexity and physiological relevance; - Tumor-like nature may introduce bias in the results obtained; - Possibility of flat surface interference in cells' polarization. - Relative incompatibility with certain food components; - Variability in transporters' expression may impair comparisons of permeability rates.
	3D (organoids)	<ul style="list-style-type: none"> - Allows cell changes in their shape; - Promotion of more complex cell-to-cell connections than other rigid systems; - Comprises all cell types and the complexity of the in vivo epithelium; - Can be cultured indefinitely; - Can be cultured from different species. 	<ul style="list-style-type: none"> - Demands a cell culture laboratory, with skilled personnel; - The established protocols are mainly focused on regenerative medicine purposes; - Expensive and demanding of complex technical skills.
	Gut-on-chip	The human Gut Chip is a microfluidic culture device composed of a transparent silicone polymer that is composed of two channels separated by a porous membrane. On one side, human intestinal epithelial cells are cultured, and human microvascular endothelial cells are on the opposite side.	<ul style="list-style-type: none"> - In comparison with organoids, gut-on-chip offers greater experimental control through multiple connected microfluidic channels; - Allows for the mimicking of the interactions of the intestine with the microbiota; - Allows for the customization of a disease phenotype.
Non-biological methods	Synthetic membranes for dialysis purposes	<ul style="list-style-type: none"> - Wide range of systems from simple to high throughputs; - Alternative for biological tissue shortages. 	<ul style="list-style-type: none"> - Does not mimic the epithelium physiology; - More sophisticated or automated systems require skills and initial investment; - Different pore sizes reported in the literature.
	Cell-free systems (PAMPA, PVPA, Permeapad® and AMI-system)	<ul style="list-style-type: none"> - Simple and ready to use, not requiring lengthy and expensive preparation steps, as in cell-based methods; - Distinct types of membranes can be applied according to the physicochemical properties of the compound in study. 	<ul style="list-style-type: none"> - Only applicable for predicting passive transcellular drug transport, since paracellular and active transports cannot be mimicked.

TEER—Transepithelial Electrical Resistance; PAMPA—Parallel Artificial Membrane Permeation Assay; PVPA—Phospholipid Vesicle-based Permeation Assay; AMI—Artificial Membrane Insert. In text references.

5.1. Tissue-Based Methods

Ex vivo methods are commonly used to predict intestinal absorption, utilizing viable and functional tissues isolated from organisms properly incubated under controlled conditions. These methods include the Ussing chamber, everted sac, and Franz diffusion cells, among others (Figure 3) Nevertheless, these ex vivo techniques imply the loss of blood flow and nervous systems, which may impair the reliability of some results. Despite this, they constitute more attainable and practical alternatives to in vivo studies [12,67].

5.1.1. Ussing Chamber

A Ussing chamber assay represents the most common technique among the ex vivo models to assess intestinal absorption. Since it was created in the 1950s to investigate sodium uptake, this system continues to be relevant nowadays for intestinal permeability studies [68]. This technique is frequently used to measure the transport of ions, nutrients, or drugs across epithelial tissues [69]. It consists of an opened intestinal segment supported, creating two isolated compartments: one corresponding to the luminal side, where the

molecule of interest is placed, and the serosal side, corresponding to the basolateral region. This system also controls and maintains temperature and continuously gasses the chamber. Moreover, two voltage-sensing electrodes and two current-passing electrodes are placed on each side, which monitor and provide current, enabling the measurement of transepithelial electrical resistance (TEER) and the short-circuit current (Isc), as a structural integrity criterion [69,70]. On one hand, the Ussing chamber technique is well validated and can be used to study permeability across distinct intestinal regions, as it allows the assessment of carrier-mediated transport and works with drugs that are poorly absorbed. On the other hand, it requires the removal of the muscular layer since contractions might interfere with electrical monitoring [71]. Moreover, not all animal models are suitable for this technique, for instance, rabbit tissues, since they are considered too thick to allow diffusion processes. Other disadvantages are the low to medium throughput of this technique, the high cost of the system and the technical skills needed. More recently, modular Ussing chamber systems have been developed and commercialized to increase the throughput, such as NaviCyte [72]. Furthermore, Westerhout et al., 2014 [73] optimized the use of porcine intestinal segments in a newly developed InTESTine™ system to predict intestinal absorption, for drug development or digested foodstuffs.

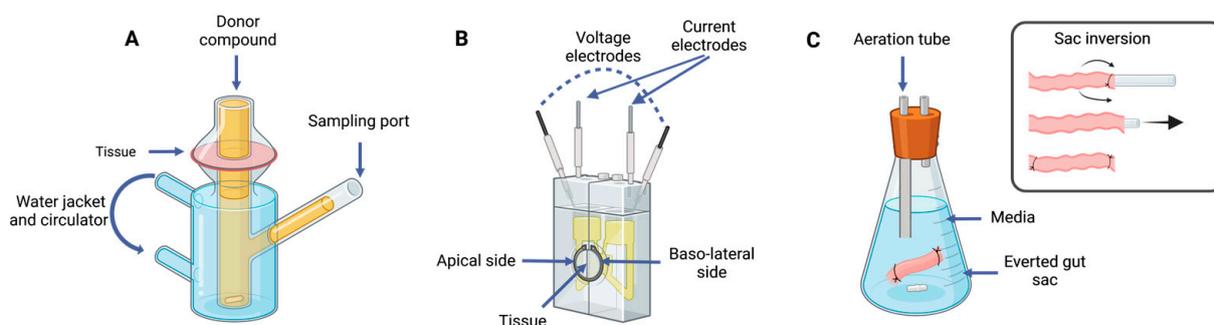


Figure 3. Tissue-based methods: (A) Franz cells; (B) Ussing chamber; (C) everted gut sac technique coupled with sac inversion diagram. Image created with BioRender.com.

5.1.2. Franz Diffusion Cells

Franz diffusion cells constitute an established system for transdermal diffusion experiments in skin tissues, as recently reviewed by Supe et al., 2021 [74]. In the literature, few studies regarding intestinal absorption rely on this apparatus. While the principle behind this system is very similar to that of the Ussing chamber, some differences are worth mentioning. For instance, Franz cells follow a vertical orientation instead of a horizontal one (as in Ussing chambers). The compartments have different volumes: the donor chamber, which receives the compound of study, has a smaller volume than the receiving chamber [13]. Moreover, there are no electrodes or gasification, but only controlled temperatures and the stirring of the receiving chamber, which may be the reason for the higher permeability when comparing with the Ussing chamber, as proposed by Dezani et al., 2013 [75].

5.1.3. Intestinal Rings and Segments

These techniques consist of exposing intestinal segments or rings, to the compound of interest, dissolved in a buffer, to assess its uptake by enterocytes and subsequent metabolism [12]. An intestinal segment is isolated and, after washing, is cut either into small rings or segments and submerged in a proper oxygenated medium. The intestinal rings might also be incubated in an everted position, as in the evert sac technique. While in the intestinal segments, the muscle layer is removed; in the intestinal rings, no layer is removed, which influences their viable time (1 h for intestinal rings and 2 h for segments). The main advantages of these methods are their practicality and simplicity, the ability to choose distinct intestinal regions and their high throughput. Additionally, one single animal can provide various rings from each intestinal region, which might be advantageous

in case of a shortage of tissues [12]. The main constraints consist of the impossibility of discriminating the direction of the transepithelial transport since both sides—the serosal and mucosal surfaces—are exposed to the compound of interest and the tissue has reduced viability over time [13,67]. Additionally, the presence of all tissue layers impairs proper oxygenation. Given these limitations, intestinal rings or segments are becoming less relevant, with few recent research papers mentioning them.

5.1.4. Everted Gut Sac

The everted sac technique was created in the 1950s by Wilson et al., 1954 [76]. In this technique, an intestinal section is reverted, and both ends are tied, after filling it with an appropriate buffer solution. Then, this intestinal section is incubated with the chosen buffer and the compound of interest. The incubation usually involves a controlled temperature and aeration or stirring. The main advantages of this method are the possibility to test distinct intestinal regions and the small volume inside the sac leading to a sample concentration, which is better in terms of analytical analysis. Additionally, it is low-cost and practical, since several samples can be tested. Furthermore, it provides an increased absorption area with a mucosal surface. However, the major drawback relies on the tissue integrity, namely the damage in the eversion process and the viability during the assay. Luo et al., 2013 [67] reported changes after only 5 min of incubation, although Verhoeckx et al., 2015 [12] described the tissue as viable for up to 2 h. Moreover, the absorption time might be slower due to the presence of all layers of the intestinal wall, which may result in underestimating the absorption. From a different standpoint, several variations to this method are worth mentioning, namely not everting the intestine segment, or instead of tying both ends, inserting a cannula in the serosal side. The everted sac method has been used to tackle the mechanisms of carrier-mediated absorption and kinetics of drug absorption and even drug metabolism. In the literature, the use of different animal tissues, namely chickens and pigs, has been reported [77].

5.2. Cell-Based Methods

Cell-based methods can be divided according to the number of cell lines involved (monocultures or co-cultures) or the growth dimensions (2D or 3D), as represented in Figure 4. Additionally, the cell lines and their origin deeply influence the quality of the models generated. The use of Caco-2 monolayers (a cell line originally derived from a colon adenocarcinoma) is frequently used for permeability and mechanistic studies regarding the intestinal epithelium [78]. This line has been widely acknowledged for its morphological and physiological resemblance with small intestine enterocytes. They can differentiate into polar columnar absorptive cells with brush borders, tight junction expressions, similar enzymes and carrier-mediated transport systems [79]. As such, *in vitro* protocols have been developed, namely the use of plate inserts. Briefly, these protocols consist of growing cells in a permeable support in a plate and allowing them to properly differentiate for about 21 days. Then, the apical and basal sides are equilibrated using a proper buffer (e.g., Hank's Balanced Salt Solution) and the formulation of interest is added to the apical side. Samples are collected from the basal side over time, and the volumes collected are balanced with a buffer. Additionally, the epithelial barrier function is monitored by measuring TEER values, before and during the assay [79,80]. While these assays with Caco-2 monolayers might be simple and practical, they fail to mimic the complexity of the intestinal environment, as they lack the diversity of cell types found in the epithelium and their interplay. For instance, goblet cells are found dispersed among enterocytes and produce the mucus that covers the whole epithelium, contributing to the barrier function and therefore affecting cellular uptake. Given these disadvantages of monocultures, nowadays, it is common to find in the literature protocols comprising two cell lines: Caco-2 and HT29-MTX [81,82]. This last cell line is also isolated from a human colon adenocarcinoma and upon treatment with methotrexate, expresses mucins, a component of mucus and differentiated into mature goblet cells. However, these cells do not express much TJs as Caco-2 does. Therefore,

Caco-2 cells are more often used alone compared with HT29-MTX, which, in turn, are commonly used in co-cultures with Caco-2. These combined cell lines are used in protocols as described earlier but require an initial step of mixing both cells in proportion, before seeding them [83]. Thus, a more complex and realistic model is obtained, with fully differentiated enterocytes and a mucus layer.

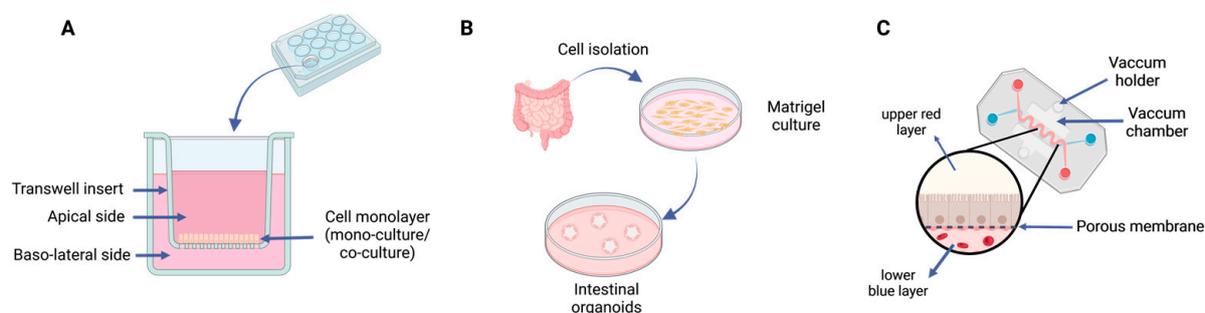


Figure 4. Schematic representation of cell-based methods for assessing intestinal absorption: (A) 2D cell culture system, either monoculture or co-culture, using an insert; (B) development of an intestinal organoid; (C) gut-on-chip technology. Image created with BioRender.com.

While the models described earlier represent the most conventional approach based on 2D models, new strategies like 3D organoids or gut-on-chip technologies, are currently being developed and optimized. The 2D models, based on growing cells on flat surfaces, fail to represent the physiological and structural *in vivo* complexity, namely regarding cell-to-cell and cell-to-extracellular environment interactions. In turn, these interactions are responsible for cell differentiation, vitality and proliferation. Additionally, because of culturing, there is an accumulation of genetic changes through time, which induces a loss of a diverse phenotype [84]. In opposition, the development of 3D organoids leads to more accurate physiological models compared with the naturally occurring structure. These models are usually created by culturing intestinal stem cells, isolated from crypt cells [12,85,86], which then communicate and differentiate efficiently through the matrix into so-called “mini-guts”. Several cell types, usually found in the epithelium, such as goblet cells, enterocytes, Paneth Cells and enteroendocrine cells can be found. Some disadvantages of this approach involve the demanding technical skills required to carry out this laborious technique. Moreover, it still does not reproduce the immune, vascular, lymphatic and enteric nervous systems, as can be done *in vivo* [12,87]. While this kind of approach has begun to be considered for drug development and regenerative medicine purposes, it has not yet been established for permeability and interaction studies with food ingredients, despite its relevance. Additionally, gut-on-chip systems are microfluidic culture devices composed of a transparent silicone polymer that is made of two channels, separated by a porous membrane. On one side, human intestinal epithelial cells are cultured, and on the opposite side, there are human microvascular endothelial cells. Therefore, this high-resolution technique allows the control of parameters such as concentration gradients, tissue–organ interactions and cell patterning, among others [88]. This system has been improved to also include stable communities of microorganisms of the gut microbiota, which allows a better understanding of host–microbiome interactions, opening a path for the development of new therapeutics or nutraceuticals [89]. This system ensures a physiologically appropriate gut environment and allows the continuous collection of fluids that flow through the lumen of the intestinal chip. The samples collected can be used for the quantification of nutrient digestion, mucus secretion or even the assessment of the status of the intestinal barrier. Gut-on-chip systems represent a valuable tool not only for pharmacological and host–microbiome studies, but also for personalized medicine, and studies on nutrition and metabolism. However, as Xiang et al., 2020 [90] pointed out, while gut-on-chip systems might be very useful for nutraceutical science, as the effects of foods on health and nutrition-related diseases might be properly evaluated, the complexity of food

represents a challenge, given the barriers in sample processing and analysis. Moreover, in general, organ-on chips have a low throughput, which can be useful for the late stages of the development/validation pipeline, but not in the early stages (screenings). Therefore, the development of high-throughput organ chips, such as on a single device with several chambers, would be highly valuable [91].

5.3. Nonbiological Methods

Another common strategy is to simulate intestinal absorption using semi-permeable membranes for dialysis purposes. The dialysis process, in general, depends on a concentration gradient as a driving force for molecule movement across the membrane and can be classified into positive or negative dialysis. In positive dialysis, the solution with the compound of interest is placed inside the dialysis bag, which, in turn, is surrounded by artificial intestinal fluid. In the reverse dialysis method, the dialysis bag is filled with intestinal fluid, and the bag is placed in a container with the compound of interest dissolved [92]. One critical factor in this technique is the pore size of the membrane, which is usually under 20 kDa [93–95]. From a different perspective, two approaches can be found in the literature regarding this process. Some authors consider dialysis a technique for simulating absorption in the small intestine, regarding it as step in BA determination. Others consider it a final step of the BC assessment, as a way to separate soluble and insoluble fractions, since only the soluble fraction can be eventually absorbed. For instance, Gayoso et al., 2016 [96] compared dialysis to centrifugation to determine the BC of food and extracts given their solubility; Moreno-Montoro et al., 2018 [93] used a combined strategy of centrifugation followed by dialysis to select which portion of the soluble fraction (centrifugation) supernatant would be absorbed (dialysate). On the other hand, it is also possible to find some standardized and complex systems for BC prediction for both drugs and foods, such as the TNO Gastrointestinal Model (TIM) or Engineered Stomach and Small Intestine (ESIN), which include dialysis membranes as a last digestion step for simulating intestinal absorption, namely passive absorption [12,97]. For example, González et al., 2020 [95] reported an improved dialysis system based on a continuous flow of fluid at a controlled temperature for a more realistic dynamic small intestine simulation. In 2007, another dialysis system arose: the AMI system. This system consists of a regenerated cellulose membrane with a molecular cutoff of 2 kDa, which is assembled between two rings. After proper validation through correlation tests with other techniques, this method established itself as a high-throughput and cost-effective tool for assessing the passive permeability of poor water-soluble compounds [52]. As such, dialysis itself represents a simple technique, accessible, but open to different approaches and improvements, given the heterogeneity of methodologies found in the literature.

Another approach for investigating intestinal absorption consists of artificial membranes, which mimic the compound's interactions with the phospholipidic cell membrane, using nanomaterials or 3D printed templates with biomolecules [98]. These new technologies might represent an interesting tissue and cell-free alternative for mimicking intestinal absorption. For instance, Permeapad™ represents an innovative membrane that can be used in the replacement of ex vivo tissues in Franz diffusion cells or as an alternative to cell culture methods, as plate inserts [99]. Additionally, PAMPA systems have also been acknowledged for having a good correlation with Caco-2 permeation assays, although failing to mimic paracellular transport. This system consists of a supported filter and a membrane soaked in phospholipids dissolved in an organic solvent and is usually applied into a 96-well plate, separated into a donor and acceptor cassette. Through time, this technique has evolved to distinct support filters, membrane compositions, solvents and pH values in the acceptor and donor chambers. Another technique worth mentioning is PVPA, in which a liposome solution is deposited in a filter support (Transwell inserts, for example). These liposomes are considered building blocks resembling the phospholipidic bilayer. This technique presents a good correlation with absorbed fraction values found in the literature for Caco-2 models and PAMPA models [52].

Considering the limitations of the techniques reviewed above, it is worth mentioning the importance of carrying out these methodologies in combination with others. For instance, it is common to combine a Ussing chamber, Franz diffusion cells or everted gut sac techniques with further validation through cell culture-based methods, generally using Caco-2 cell lines for Transwell® assays. For instance, Westerhout et al., 2014 [73] developed a new approach to assess intestinal permeation and further validated the results of several drug permeations through a comparison with a co-culture of Caco-2 and HT29-MTX on a Transwell® system. B Sánchez et al., 2019 [100] also tested different drugs using Franz cell diffusion assays with porcine tissues, and the results were compared to the results found in the literature regarding permeation values obtained with Caco-2. Chen et al., 2019 [101] tested the permeation of chitobiose using an everted rat gut sac and compared it with the results of a Transwell® assay with Caco-2. While these examples refer to technique validation and drug permeability purposes, this approach could constitute a good practice to implement in biomolecule/ingredient development studies.

Table 2 summarizes the literature review regarding the application of the methodologies previously described, for ingredient/formulation development. It is possible to point out a clear gap in approaches that favor the *in vitro* pre-assessments of ingredients' potential, especially concerning chicken models. From the literature review performed, it was possible to notice that most studies report only digestibility results, by simulating enzymatic GIT, while failing to consider intestinal absorption; other studies tackled the effects of some ingredients on intestinal permeability, either directly, with no prior digestion, or after their *in vivo* administration. Overall, studies on promising ingredients, reporting both BC and BA to some extent (GIT simulation and intestinal absorption), are scarce or inexistent. The outlook on *in vitro* studies looking toward human health is much more positive as studies including *in vitro* digestion simulation followed by intestinal absorption prediction could be found. Moreover, porcine models provide an opportunity for *ex vivo* studies on intestinal absorption. Cell culture models rely on well-established protocols, using human-derived cell lines. While for chicken feed, no cell culture protocol has been used for ingredient development, and only a few exploratory studies with primary cell lines or organoid development could be found [102].

Table 2. Review of approaches found in the literature used to assess the intestinal permeation of functional compounds or ingredients.

Technique	Food/Ingredient Tested	Model	Description	Reference
Ussing chamber	Cinnamon bark oil and coconut oil emulsions	Laying hen	No previous simulation of the gastrointestinal tract	[103]
	Apple polyphenols	Pig	Direct application of polyphenols to a Ussing chamber; no previous digestion of the polyphenols was performed	[104]
	Organic acid supplementation in feed	Chicken	<i>In vivo</i> administration of the supplement was performed; a Ussing chamber was used to assess intestinal permeability changes	[105]
	Oligopeptides from whey protein hydrolysate	Pig	<i>In vitro</i> digestion and absorption simulation through Ussing chamber;	[106]
Franz cells	Tetrahydrocurcumin-hyaluronic acid conjugate (metabolite of curcumin)	Pig	TNO dynamic gastrointestinal model-1 (TIM-1) followed by Franz cell assay	[107]
	Monosaccharides, amino acids and a corn oil-in-water emulsion	Semi-permeable cellulose membrane	No gastrointestinal simulation was performed; no biological tissue was used	[108]

Table 2. Cont.

Technique	Food/Ingredient Tested	Model	Description	Reference
Everted intestinal sac	Fructose uptake	Chicken	No previous digestive process was simulated	[109]
	Garra fish meal	Chicken	In vivo studies carried out in chickens, followed by an evaluation of diet effects on intestinal permeability	[110]
	Encapsulation of β -carotene in zein protein	Chicken	Study focused on human health; human GIT digestion simulation was simulated, followed by absorption experiments with chicken intestines	[111]
	Phenolic compounds from non-extruded and extruded Mango Bagasse-added confections	Pig	Human in vitro GIT digestion, followed by permeability assessment	[112]
	Encapsulated curcumin and resveratrol	Pig	Human in vitro GIT digestion was carried out, followed by an everted gut sac for BA assessment	[113]
Mono-cultures	Angiotensin I-converting enzyme inhibitory peptides, from cooked chicken breast/thighs	Caco-2	Peptides identified after in vitro digestion, followed by PET inserts with a Caco-2 monoculture	[114]
	Curcumin alone or with polyvinylpyrrolidone	Caco-2	This study focused on chickens' health; despite an in vitro simulation of the chicken GIT, a Transwell permeability assay was performed with the initial samples, not the digested ones	[115]
Co-cultures	Sardine protein hydrolysate	Caco-2 + HT29-MTX	After human in vitro digestion, permeability was assessed using PET inserts (like the Transwell™ system)	[116]
	Encapsulated rosemary extract	Caco-2 + HT29-MTX	In vitro human digestion followed by co-culture in a Transwell™ system	[117]
	Salmosan (derived from Mannan oligosaccharide)	Caco-2 + THP-1 (macrophages)	Salmosan and Salmosan with <i>L. plantarum</i> were tested for the effects on the intestinal permeability and barrier, as potential feed additives; nevertheless, no GIT digestion was simulated	[118]
Dialysis membrane	Gelatinized starch dispersions	Hollow fiber membrane (synthetic)	Study on starch digestion and the consequent absorption of hydrolytic products generated in the human small intestine, using an in vitro intestinal digestion system (i-IDS)	[95]
	Phenolics, flavonoids, rutin, β -carotene and lutein in six edible greens	Cellulose membrane (12,000 Da)	Bioaccessibility and BA was evaluated; GIT simulation was carried out followed by dialysis, as a simplified model of intestinal permeation	[119]

Table 2. Cont.

Technique	Food/Ingredient Tested	Model	Description	Reference
PAMPA	Crude plant extracts (<i>Angelica archangelica</i> , <i>Waltheria indica</i> , <i>Pueraria montana</i> var. <i>lobata</i>)	Polycarbonate filter plate (5–20% porosity with a 0.45 µm pore size and 9–10 µm thickness) impregnated with a hexadecane/hexane (5/95 % (v/v)) solution	Prediction of the passive intestinal absorption of a representative set of frequently occurring natural products from <i>Angelica archangelica</i> , <i>Waltheria indica</i> and <i>Pueraria montana</i> var. <i>lobata</i> ; no GIT digestion simulation was performed before PAMPA assay	[120]
	Saponins and sapogenins from seed extracts from red quinoa and seeds of fenugreek.	Lipid mixture containing L- α -phosphatidylcholine and cholesterol in 1,7-octadiene solution added to the PVDF filter of each well; after membrane coating, the donor solutions were added	Evaluation and comparison of the permeability of saponins and sapogenins from fenugreek and quinoa extracts with and without previous in vitro digestion simulation, through the previous development of a GIT digestion protocol attached to PAMPA	[121]

The literature revision, according to the defined criteria, underlines the lack of relevance of intestinal segments or of the ring technique since no research papers were found on these topics in the period considered. Moreover, the reduced number of Franz diffusion cell studies is indicative of the reduced applicability of intestinal permeability assays. Few papers are designed in a way to fully assess the influence of the gastrointestinal tract. For instance, most do not simulate the process from mouth to colon, neglecting this relevant topic. As for chickens, there is a relevant lack of cell culture-based methods and non-biological methods for feed/nutritional studies.

6. Conclusions and Opportunities

Overall, studies regarding physiology aspects and nutrient transport mechanisms for porcine and/or human models are more abundant than for chicken models. The same trend is observed for studies focused on the development of functional ingredients or bioactive compounds that include the GIT and intestinal absorption simulation.

The study of the impact of new ingredients and molecules on human and animal health demands reliable and complete evaluation methods, from mouth to colon, preferably including potential interactions with the microbiota. Concerning in vitro models for predicting bioaccessibility, there are several models, either static or dynamic, but also simulating one digestion compartment or several in sequence. Some models are more suited for mechanistic studies, while others are more adequate for kinetic studies. Overall, INFOGEST represents an adequate model for either the kinetics of nutrient release or for monitoring the concentration or stability of a particular substance or nutrient. Moreover, it is easy to implement and reproduce and allows comparisons between distinct laboratories and studies pointing to a good correlation with in vivo studies. However, the same broad consensus was not attained for animal digestion simulation models or concerning models that assess the potential interactions with the microbiota. Additionally, for both humans and animals, the absorption simulation step lacks standardization and consensus among the scientific community. The studies using absorption models for chickens' nutrition and feedstuff development were overall scarce. Currently, there is no "one size fits all" technique for simulating intestinal permeation. Different techniques require distinct implementation costs and technical skills and present distinct pros and cons. Therefore, the strategies employed in some drug-related studies combining the use of two techniques—for instance, an ex vivo method along with a cell-based method—provide more reliable results and should constitute a general recommendation in the nutritional studies field. Moreover, in drug development studies regarding solubility and absorption, these follow highly standardized classification systems and testing protocols (for example, the Biopharmaceutical Classification System (BCS)). In fact, even dissolution in nutritional liquids has been explored [122]. The guidelines used in toxicology might be useful for reflecting on and in

the application of the study of functional ingredients' biopotential. Additionally, the need for complete GIT simulation models constitutes an important step before in vivo studies, in alignment with widespread ethical standards.

In summary, the purpose of this review was to bring awareness to the lack of complete standardized models—from mouth to colon—needed to achieve meaningful conclusions when studying the potential of bioactive ingredients, either for human or animal applications. Our recommendation for reliable in vitro screenings when developing new ingredients/formulations for food or feedstuffs would be to adopt an in vitro gastrointestinal digestion strategy: a model that is as standardized as possible with colonic/cecal fermentation followed by a cell culture-based method (a Transwell assay, for instance) with an ex vivo tissue-based model before laborious and expensive in vivo testing. Nevertheless, this general approach would benefit from a proper in vivo study to establish a correlation between them.

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